

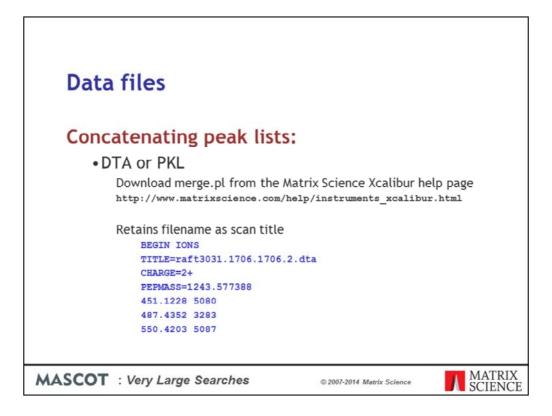
Very large searches present a number of challenges. These are the topics we will cover during this presentation.

	۲	Mascot Daemon	
• Can use Mascot Daemon to process and merge MudPIT	Ele Edit Help Status   Event Log Task Estor   Pa Owner Parameter set   human Oata file list	Task Munungous search New Data import filter Istacord Datalar  Schedule	Run Options
fractions • Use Distiller or a file specific data import filter	Drog and flore data Blen in before or clink on Add 4 Gample TUD 06222014, sa 4 Gample TUD 0622014, sa 4 Gample TUD 0620014, sa 4 G	C Start at     TSeptember2014     C Real-Sene monitor     C Follow-up     C Follow-up     C Follow-up     Auto-export     Auto-export     Lose	

The smartest way to merge files, like fractions from a MudPIT run, is using Mascot Daemon. Just tick the box at the bottom left.

The batch can be peak lists or raw files

Note that Mascot Daemon 2.1 had a file size limit of 2 GB. This was lifted in 2.2, and we have successfully merged and searched a 6 GB file, although note that some web servers cannot accept uploads larger than 4 GB



If you don't want to use Daemon, you can merge peak lists manually.

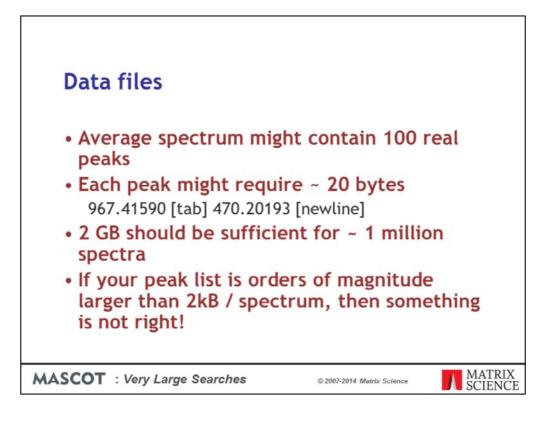
For DTA or PKL, you can download a script from our web site.

A nice feature of this script is that it puts the filename into the scan title, so you can tell which fraction a particular spectrum came from. The scan titles are displayed in the yellow pop-ups on the Mascot result report

Data files	
•MGF	k lists:
Windows: copy	C:\TEMP>copy *.ngf merge.out
Unix: cat	<pre></pre>
MASCOT : Very Large Searche	es © 2007-2014 Matrix Science MATRIX SCIENCE

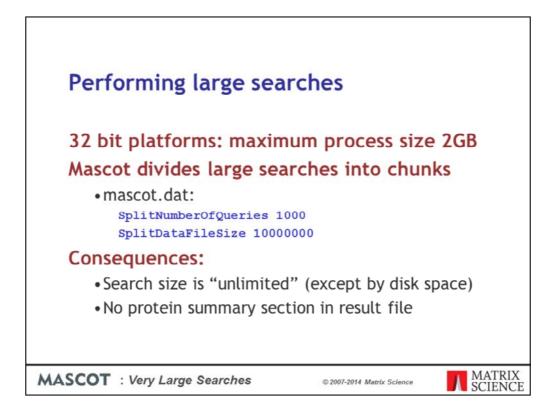
As long as MGF files contain only peak lists, you don't need a script. Just use copy or cat

If the MGF files have search parameters at the beginning, you'll need to remove these before merging the files. Because a number of third party utilities add commands to MGF headers, and these cause a merged search to fail, Mascot Daemon 2.3 and later strips out header lines when merging MGF files.



In talking to Mascot users, it is clear that peak lists files are often much bigger than they should be. In other words, the peak detection is not very good. If you do a back of the envelope calculation, you can see that 2 GB should be enough for approximately 1 million spectra.

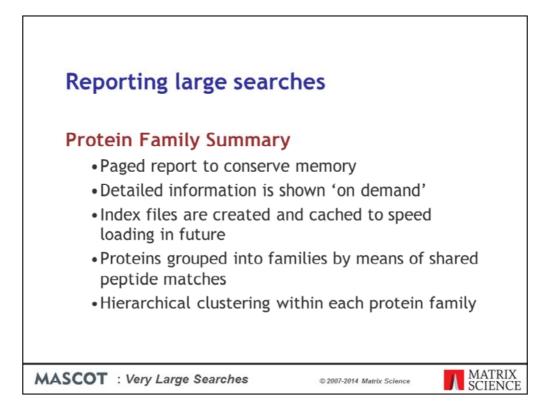
If you intend to do a lot of large searches, its worth getting the peak detection right. Shipping unnecessarily large files around wastes both time and disk space



32 bit platforms have a maximum process size of 2 GB on Windows or 3Gb on Linux. To get around this limit, Mascot divides large searches into smaller chunks, so as to avoid having everything in memory at the same time. The parameters to control this are SplitNumberOfQueries and SplitDataFileSize in the Options section of mascot.dat

One consequence of splitting a search is that there is no protein summary section in the result file. This is not a problem, because no-one wants a protein summary report for a large MS/MS search. However, some old client software gets confused by the missing section. The work around is to increase the values so that large searches never split. Maybe setting SplitNumberOfQueries to 1 million spectra and SplitDataFileSize to 10 billion bytes.

This is OK, but remember to reset these values as soon as you are able to. Otherwise, you might find you run out of memory or address space for your large searches



In early versions of Mascot, trying to display result reports for very large searches would often lead to problems with timeouts and running out of memory. To address this, the Protein Family Summary loads most of the information 'on demand'. This requires some index files to be created on the server, and these index files are cached, so that the report loads much faster on the second and subsequent occasions. Proteins are grouped into families by means of shared peptide matches and, within each family, hierarchical clustering is used to illustrate which proteins are closely related and which are more distant.

C 54.243.190.62/m	nascot/cgi/master_results_2.pl?file=_%	2Fdata%2F	F981139.dat	
eins (479) Report Build	fer Unassigned (27741)			5.pem
ein families 1-10 (o	ut of 479)			
	1 2 9 - 10 Next Expand al	Collapse		
		Compt		
sion • contains •			Find	
	1::sp[TRY1_BOVIN]	1606	sp[TRV1_BOV3N]	
	1 2::CP2CT_MOUSE	1337	Cytochrome P450 2C29 OS=Mus musculus GN=Cyp2c29 PE=1 SV=2	
	6 2::CP239_MOUSE	252	Cytechrome P450 2C39 OS=Mus musculus GN=Cyp2c39 PE=2 SV=2	
	7 2::CP238_MOUSE	150	Cytochrome P450 2C38 OS+Mus musculus GN+Cyp2c38 PE+2 SV+2	
	2 2::CP254_MOUSE 8 2::CP270_MOUSE	553 73	Cytochrome P450 2C54 OS=Mus musculus ON=Cyp2c54 PE=2 SV=1 Cytochrome P450 2C70 OS=Mus musculus ON=Cyp2c70 PE=2 SV=2	
	3 2::CY250_MOUSE	490	Cytochrame P450 2C50 OS=Mus musculus GN=Cyp2c50 PE=1 SV=2	
_	5 2::CP237_MOUSE	339	Cytochrame P450 2C37 OS=Mus musculus GN=Cyp2c37 PE=2 SV=2	
	4 2::CP2F2_MOUSE	485	Cytochrome P450 2F2 OS+Mus musculus GN=Cyp2f2 PE=2 SV=1	
200	0			
		1200		
-	1 2::GRP78_MOUSE 2 2::HSP7C_MOUSE	362	70 kDa glucose-regulated protein OS=Mus musculus GN=Hspa5 PE=1 SV=3 Heat shock cognate 71 kDa protein OS=Mus musculus GN=Hspa8 PE=1 SV=1	
L	3 2::HS71L_MOUSE	188	Heat shock 70 kDa protein 1-like OS+Mus musculus ON+Hapa1I PE+2 SV+4	
<u> </u>	z •			
	2::CYB5_MOUSE	1217	Cytochrome b5 OS=Mus musculus ON=Cyb5a PE=1 SV=2	
	2::PDIA1_MOUSE	1124	Protein disulfide-isomerase OS=Mus musculus ON=P4hb PE=1 SV=2	
	2::CP1A2 MOUSE	1054	Cytochrome P450 1A2 OS+Mus musculus GN+Cyp1a2 PE+1 SV+1	
	2::ENPL_MOUSE	1018	Endoplasmin OS=Mus musculus ON=Hsp90b1 PE=1 SV=2	
-	1 2::RDH7_MOUSE 2 2::H1786_MOUSE		Retinol dehydrogenase 7 OS+Mus musculus ON+Rdh7 PE+2 SV+1 17-beta-hydroxysterold dehydrogenase type 6 OS+Mus musculus ON+Hud1765 PE+2 SV+1	
11111	T Introduction	200		
ਬੰ ਸੱ ਵੱ ਸੱ ਸੱ	ਜੋ ਛੋ			
	2::MGST1_MOUSE	863	Microsomal glutathione S-transferase 1 OS+Mus musculus GN+Mgst1 PE+1 SV+3	
	2::RL7A_MOUSE	770	605 ribosomal protein L7a OS=Mus musculus GN=Ral7a PE=2 SV=2	

If there are 300 or more spectra, the Protein Family Summary is the default. This is the appearance of a typical family report immediately after loading. The body of the report consists of three tabs, one for protein families, one for Report Builder, and one for unassigned matches. The report is paged, with a default page size of 10 families. If you wish, you can choose to display a larger number of families on a single page.

Proteins are grouped into families using a novel hierarchical clustering algorithm. If the family contains a single member, the accession string, protein score and description are listed. If the family contains multiple members, the accessions, scores and descriptions are aligned with a dendrogram, which illustrates the degree of similarity between members.

The scores for the proteins in family 2 vary from 1337 down to 73. In the earlier Peptide Summary or Select Summary reports, these would have been at opposite ends of the report. It would have been difficult to recognise that these proteins belonged together, even though they have shared peptide matches and are all cytochrome P450.

	54.243.1	90.62/mas	cot/cgi/mas	ter_results_	2.pl?file=_%2	2Fdata%	2FF9811	39.dat	jgnoreionsscorebe	low=0.05:_prefertaxonomy=0:_sigthreshold=0.05:percolate=i {	2 6	1
				ECP2CT_M						us GN+Cvp2c29 PE+1 SV+2		
				::CP239_M		25				us GN=Cyp2c39 PE=2 SV=2		
T I				CP238_M		13				us GN+Cyp2c30 PE+2 SV+2 us GN+Cyp2c54 PE+2 SV+1		
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1				::CP237_M		33				us GN+Cyp2c37 PE+2 SV+2		
-			4 2	CP2F2_M	OUSE	41	4 Cytoch	Irome P4	150 2F2 OS+Mus musculut	GN+Cyp2f2 PE+2 \$V+1		
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			Score	Mass		Sequen		PAI				
2.1	d2::CP2CT_		1337	61419	76 (76)	13 (	2018 I			+Hus musculus GN=Cyp2c29 PE+1 SV=2		
2.2	#21:CP254_	MOUSE	552	60887	27 (27)		4-4	0.88 0	Sytechrome P450 2C54 OS	Hus musculus GN=Cyp2c34 PE=2 SV=1		
2.3	#2::CY250_	MOUSE	489	61128	27 (27)	10 (	10)	1.20 0	Cytochrome P455 2C50 OS	+Mus musculus GN=Cyp2c50 PE+1 SV+2		
2.4	#2::CP2F2_	MOUSE	484	59267	32 (32)	12 (	12)	2.11	Cytachrome P450 2F2 05+	Mus musculus QN=Cyp2f2 PE=2 SV=1		
2.5	#21:CP237_	MOUSE	339	60590	22 (22)	8	(8)	0.89	Cytechrome P450 2C37 OS	+Hus musculus GN+Cyp2c37 PE+2 SV+2		
2.6	#2::CP239_	MOUSE	251	60856	13 (13)	4	(4)	0.37	ytechrome P450 2C39 OS	+Mus musculus GN+Cyp2c29 PE+2 SV+2		
6 2.7	#2::CP238_	MOUSE	150	61356	9 (9)	4	(4)	0.37	Cytechrome P450 2C38 OS	+Mus musculus GN=Cyp2c38 PE+2 SV=2		
Redispla	All Non											
Auto-f	<b>tide matches (</b> lit to window	43 non-dup										
12346	Y Dopes		Mr(expt) 1004.6178		Delta M 0.1095 0		Expect 0.015		0 1 2 3 4 5 6 7	R.MPTLEDR.T		ź
			1005.7547				0.014		0	R.FSVQILR.N	1	Î
ef419			1031.7808				0.049			VOREIDR.V		
	7 12		1560.7029				1.2e-05		0	K.NISQSETNESR.A		١
		521.3753	1040.7361	1040.5810	0.1551 0	22	0.031			R.FTLMTLR.N + Oxidation (M)		
	4		1573.3479	1572.7654	0.5824 0	71	1.3e-05			K. EALVDHOEEFAOR. O		
£444				1050.5323	0.0453 0	35	0.0084			R.CLVEBLR.K		
18444 18446 18470			1050.5776			54	0.00019			R. ICAGEGLAR.M		
ದೆ 444 ದೆ 446 ದೆ 470 ದೆ 973	5	526.2961	1050.5776	1078.5385	0.0964 0							
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ದೆ 444 ದೆ 446 ದೆ 470 ದೆ 473 ದೆ 554 ದೆ 560	5 1 <b>)</b> 3 6 <b>)</b> 8	526.2961 540.3247 541.3848	1078.6349	1080.6059	0.1492 0			11		R. YEOVTAR. V B. OSFIMAER. I		
ದೆ 444 ದೆ 446 ದೆ 470 ದೆ 473 ದೆ 554 ದೆ 560	5 L > 3 H > 8 S > 5 I > 5	526.2961 540.3247 541.3848 577.9297	1078.6349 1080.7551	1080.6059 1153.6045	0.1492 0	53 49	0.00034	11 11				

If you are interested in family 2, then you click to expand it to show the details. Immediately under the dendrogram is a list of the proteins. The table of peptide matches is similar to that found in the other result reports. Duplicate matches to the same sequence are collapsed into a single row. The columns headed 1, 2, 3, etc. represent the proteins and contain a black square if the peptide is found in the protein. Some matches are shared, but each protein has some unique peptide matches, otherwise it would be dropped as a sub-set.

In this screen shot and the ones that follow, we've set an expect cut-off of 0.05 to simplify the picture by removing low scoring matches

	0 54.243	.190.62/mas		ter_results_		2Fdata%				ssscorebelow=0.05;_prefertaxonomy=0;_sigthreshold=0.05;percolate=1 $c_2^{(2)}$	Ð
-				HSP7C_M						protein OS+Mus musculus GN+Hspaß PE+1 SV+1	
L				::HS711_M						-like OS+Mus musculus GN=Hspa11 PE=2 SV=4	
4	5 <u>8</u> =	3 ¥ X	•								
Th	reshold (0):	0	Cut								
3.1	#2::GRP7	8 MOUSE	5core 1308	Mass 81404	Matches 55 (55)	Sequen 21 (		2.47 1	to kDa gluces	r-regulated protein OS+Mus musculus GN+Hspa5 PE+1 SV+3	
3.2	#2::HSP7		362	78937	21 (21)					pate 71 kDa protein OS=Mus musculus GN=Hspaß PE=1 EV=1	
13.3	2::H571		188	78552	13 (13)					kDa protein 1-like OS+Mus musculus GN+Hspa11 PE+2 SV+4	
Redispla											
11460) 115924 116994	11	546.9979	1045.9413 1091.9813 1129.7232	1091.6430	0.3038 0 0.3383 0 0.1131 0	41	0.038	11	0	R.NTWFTK.K K.ITITNOK.G R.LTFEEIER.M	
6751			1145.9377		0.2641 0		0.0061		υ .	R. OTLDEVER. A	
ef\$021			1191.1306		0.4581 0		0.0028			R.VMERFIZ.L	
ef 9451			1205.7264		0.0517 0		5.5e-05 0.0038		0	K.VLEDSDLK.K K.ITITNDQNR.L	
1996			1220.8737		0.1872 0		0.0001			K.VCNPIITK.L	
af10031			1835.4126		0.5923 0		0.0075			K. STAODTILOGEDFDNR.M	. 1
af11545	1 C C C C C C C C C C C C C C C C C C C	635.4900	1268.9654	1268.6856	0.2799 0	55	0.00058	11		R. ETAEAYLOR. R	. 1
#1194		641.5476	1281.0806	1200.7220	0.3586 0	55	0.00015	11	0	K.EIAEAYLOR.T	. 1
W18194			1479.1791		0.3455 1	42	0.0011			K. VYEOERPLTK. D	. 1
#1819 #19454			1479.5652		0.7316 1	26	0.014			K. VYEGERPLTK. D	
			2303.9974		0.7344 1	39	0.0053 3.2e-05			K. KVTRAVYTVEAYENDAGR. Q B. NELESYAYSLK. N	
			1672.5930		0.6699 1	28	0.018			R.MRETARAYLOR.R	
#21054			1709.8639			66	2.1e-06			R. ITPSYVAFTEOER. L	
d22754					0.2770 0	57	1.78-05			R. ITPSYVAFTPEGER, L	
			1710.1517								

Moving down to family 3, the scale on the dendrogram is ions score, and HSP7C\_MOUSE and HS71L\_MOUSE join at a score of approximately 30. This represents the score of the significant matches that would have to be discarded in order to make one protein a sub-set of the other. These two proteins are much more similar to one other than to GRP78\_MOUSE, which has non-shared peptide matches with a total score of approximately 145. Note that, where there are multiple matches to the same peptide sequence, (ignoring charge state and modification state), it is the highest score for each sequence that is used.

Immediately under the dendrogram is a list of the proteins. In this example, because SwissProt has low redundancy, each family member is a single protein. In other cases, a family member will represent multiple same-set proteins. One of the proteins is chosen as the anchor protein, to be listed first, and the other same-set proteins are collapsed under a same-set heading. There is nothing special about the protein picked for the anchor position. You may have a preference for one according to taxonomy or description, but all proteins in a same-set group are indistinguishable on the basis of the peptide match evidence.

The table of peptide matches is similar to that found in the other result reports. Duplicate matches to the same sequence are collapsed into a single row. Click on the triangle to expand.

The black squares to the right show which peptides are found in which protein. To see the peptides that distinguish HSP7C\_MOUSE and HS71L\_MOUSE, clear the checkbox for GRP78\_MOUSE and choose Redisplay.

, _								-	
			::GRP78_M		1308		se-regulated protein OS=Mus musculus GN=HspaS PE=1 SV=3 Ignate 71 kDa protein OS=Mus musculus GN=HspaS PE=1 SV=1		
		3 2	HS71L_M		188		kDa protein 1-lika OS=Mus musculus ON=Hspa119E=2 SV=4		
51	8 8 8 8 8								
<		•							
Thres	hold (0): 0	Cut							
3.1	2::GRP78_MOUSE	Score 1308	Mass 81404	Matches 55 (55)	Sequence 21 (21		78 kDa glucese-regulated protein OS+Mus musculus GN+Hapa5 PE+1 SV+3		
	2::HSP7C_MOUSE	362	78937	21 (21)	8 (8		Maat shock cognete 71 kDe protein OS=Mus musculus OK=Hspeß PE=1 SV=1		
	2::HS71L MOUSE	188		13 (13)	4 (4		Meat shock 70 kDa protein 1-like OS=Mus musculus GN=Hapa1I PE=2 SV=4		
	All None			10 (10)		/			
d2720 ) d5924 ) d7529 d7529 d7529 d7529 d71946 ) d25277 d26376 d26946 d26947 2 subsets an	1 546.9979 573.9761 2 611.4441 1 612.8115 1 641.5476 607.4422 953.0936 650.1325	1091.9813 1145.9377 1220.8737 1835.4126 1281.0806 1819.3048 1904.1726 1947.3756 1947.4139	1145-6536 1220-6865 1834-8204 1280-7220 1818-8255 1903-3845 1947-0920 1947-0920	0.1363 0 0.3383 0 0.2041 0 0.1872 0 0.3586 0 0.4793 0 0.1881 0 0.2836 0 0.3218 0	41 0 38 0 60 0 35 0 55 0 55 3 84 1 37	.00024 )1 0.0061 )1 0.0061 )1 0.0001 )1 0.0001 )1 1.00015 )1 20-05 )1 0.013 )1 0.013 )1	<ul> <li>B. LIODAAR, N</li> <li>F. LITTNER, G</li> <li>B. OTLEPPER, A</li> <li>K. VUNDITFE, L</li> <li>K. STACOFILOGEDERS, M</li> <li>K. ELAZATUR, T</li> <li>R. ATAGETHLOGEDERS, L</li> <li>K. STREPSAMENTER, M</li> <li>B. IINEFFAALAINOLER</li> <li>B. IINEFFAALAINOLER</li> </ul>		
		2	::CYB5_MO	USE	1217	Cytochrome I	5 OS+Nus musculus ON+Cyb5# RE=1 SV+2		
		2	PDIA1_M	OUSE	1123	Protein diauff	de-isomerase OS+Mus musculus ON+P4Nb PE+1 SV+2		
		2	CP1A2_M	OUSE	1054	Cytochrome I	450 1A2 05=Mus musculus GN=Cyp1a2 PE=1 3V=1		
			ENPL_MO			and the second	05-Mus musculus GN-Hap9001 #E-1 SV-2		

It can now be seen that HS71L\_MOUSE would be a sub-set of HSP7C\_MOUSE if it was not for one match, K.ATAGDTHLGGEDFDNR.L. It is the significant score for this match that separates the two proteins in the dendrogram by a distance of 32 (score of 55 - homology threshold score of 23).

You can "cut" the dendrogram using the slider control.

				GRP78_M	OHEE					ted protein OS+Mus musculus GN+Hzpa5 9E+1 SV+3	9
1				HSP7C_M						LLDa protain OS+Mus musculus GN+Hspaß PE+1 SV+1	
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	5 8 B	8 8 8									
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Thr	eshold (50	): 50	Cut								
	-	a manuer	Score	Mass		Sequen		PAI			
3.1	d2::GRP7		1308	81404	55 (55)		1008			plucese-regulated protein OS=Mus musculus GN=Hapa5 PE=1 SV=3	
3.2	821:HSP7	C_MOUSE	362	78937	21 (21)		(8)	0.63	Heat sho	ick cognate 71 kDa protein OS=Mus musculus ON=Hspaß PE=1 SV=1	
Redisplay	All No	anie .									
ió peptide	e matches	(32 non-duplic	ate, 34 dup	licate)							
Auto-fit	to window										
Query	Dupes	Observed	Mr(expt)	Mr(calc)	Delta M	Score	Expect	Rank	01	2 Peptide	
#2720		488.3756	974.7367	974.6004	0.1363 0	54	0.00024		÷ 🕯	R.LIGDAAR.N	
#4601	12	523.9779	1045.9413	1045.6375	0.3038 0	33	0.038	11	U .	R.NTWFTK.K	
15924	11	546.9979	1091.9813	1091.6430	0.3383 0	41	0.0061	11		K.ITITNDK.G	- 1
ef6224		545.8689	1129.7232	1129.6101	0.1131 0	32	0.927	11		R.LTPEEIER.H	- 1
17519		573.9761	1145.9377	1145.6536	0,2041.0	38	0.0061	11	σ	R.OTLDEVER.A	- 1
#9021		596.5726	1191.1306	1190.6725	0.4581 0	45	0.0028	11	Ψ.	R. WEHFIR. L	- 1
19459	12	603.8705	1205.7264	1205.6747	0.0517 0	61	5.5e-05	11	υ .	K.VLEDSDLK.K	
19857	13	609.9429	1217.0713	1217.6486	0.2227 0	45	0.0036	11	υ 🔳	K. ITITNDQNR. L	
deeth	12	611.4441	1220.0737	1220.6865	0.1872 0	60	0.0001	¥1	U	R.VCNPIITE.L	
ef10037	11	612.0115	1035.4126	1834.8204	0.5923 0	35	0.0075	11		K. STAODTHLOGEDFONR . N	
111545		635.4900	1268.9654	1268.6856	0.2799 0	55	0.00058	11	υ .	K.ETABAYLOK.K	- 1
af11946	11	641.5476	1281.0806	1280.7220	0.3586 0	55	0.00015	11	U	K.EIAEAYLOR.T	. 1
#18194		740.5968	1479.1791	1478,8336	0.3455 1	42	0.0011	11	υ 🔳	K. VYBOERPLTK. D	. 1
10197		494.1997	1479.5652	1478.0336	0.7316 1	26	0.914	11	υ .	K. VYBGBRFLTK. D	
19656		769.0064			0.7344 1	39	0.0053	11	0 🔳	R. RYTHAVYTVIAYPHDAQR. Q	
821354	11	803.1397			0.4311 0	63	3.2e-05	11	υ .	R. NELESYAYSLE. N	
\$22784		558.5383	1672.5930	1671-9231	0.6699 1	28	0.018	11		R. MRETARAYLOR. R	
123465	>2	055.9392	1709.8639	1709.8746	-0.0108 0	66	2.1e-06	11	υ.	R. ITPSYVAFTPEGER. L	- 1
122472		571.0578	1710.1517	1709.8746	0.2770 0	57	1.7e-05	11	υ 🔳	R. ITPSYVAFTPEGER. L	
DA1112				1717.8879	-0.5253 0	77	9.4e-07			R. THERPSVOQDIK.F	

If we cut the dendrogram at a score of 50, HS71L\_MOUSE will be dropped because it is now a sub-set protein. If you compare the matches to HSP7C\_MOUSE with those to GRP78\_MOUSE, it is clear that these are very different proteins. They are part of the same family because of two shared matches, but many highly significant matches would have to be discarded for either protein to become a sub-set of the other. In summary, we can quickly deduce from the Family Summary that there is abundant evidence that both GRP78\_MOUSE and HSP7C\_MOUSE were present in the sample. There is little evidence for HS71L\_MOUSE. It is more likely that the HSP7C\_MOUSE contained a SNP or two relative to the database sequence.

	H48) Report 8	Sullder Unassig	med (30397)	7					5 permalio
Inotain fa		11							
rotem ia	milies 41-5	0 (out of 448)	1						
10 • per j	page Etexio	SIS 1 2 2 4 5	Q Z B	2 12 - 4	Next	E	xpand	all	Collapse all
Sequence	is equal to	MNVLADALK				Find	Clea	a	
						-) CONTRACTOR	ALCONT OF		
41	2::NB5R3_MO	JSE 36	54 NADH-cyto	chrome b5 reduc	tase 3 OS	+Mus musc	ulus GN	+Cyb	5/3 PE+1 SV+3
42	2::RS19_MOUS	SE 30	50 405 ribese	mal protain 519	05+Mus n	usculus GN	+ F.p.13	- 25	1 5/+3
43	2::CP2E1_MOL	ISE 35	SB Cytechreen	+ P455 2E1 05+	Hus musc	Aut Ghancy	241 PE	-2.5	V=1
	2::RL22_MOUS			mal protein L22 (					
45	2::RS15A_MOL			mal protein \$15a				28 PI	E#1 34#5
45.1	2::RS15A_MOL	JSE 344	Mass 16651	Matches : 16 (16)	Sequen		2.12	105 r	bosomal protein 515a OS+Mus musculus Olix=Rps15a PE+1 SV+2
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#13708		.3777 1014.7407		0.1100 0	45	0.00053			K. IVVNLTGR. L
ef11285 -		.9663 1261.9180 .8868 1261.7591		0.1872 0	77	2.4e-06			R. MIVLADALK . S R. FOVLADALK . S
		.8914 1261.7682		0.0375 0	(59)	9.7e-05			R. MINVLADALM . S
		.9416 1261.8686		0.1379 0	(59)	0.00013			R. HNVLADALK . S
	d11207 #32	.0080 1262.0014	1241.7308	0.2704 0	(4.2)	0.0045	1:		R MONTADALK S
	632	.0218 1262.0291	1261.7308	0.2983 0	(63)	6.4e-05	11	σ	R. NNVLADALK . S
W11604		.4751 1270.9355		0.2452 0	28	0.03			K.WOMNLLPSR.Q
#11780 ·		.8954 1277.7762		0.0505 0	50	0.00084			R. MNVLADALE.S - Oxidation (M)
	#11/79 #J9	19899 1211.9862	1211.1251	0.2396 0	(40)	0.00054	11		R. HNVLADALR. S + Omidation (M)

The family report also includes a text search facility, which is particularly important for a paged report. You can search by accession or description sub-string, or by query, mass or sequence. Here, for example, we searched for a peptide sequence. The display jumps to the first instance of the sequence, expands, and highlights (in green) the target peptides.

		49) Rep	ort Builder Unassign	ed (3039)	2						<u>&amp; permaliok</u>
roteir	ı hi	ts (476 p	proteins)								
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anning	1	CRAP	#1::sp[TRY1_BOVIN]	1606	28266	48	Hatch(sig) 48	3equences	Sequing	2.86	
	1	SwissProt	d2::CP2CT_MOUSE	1337	61419	76	76	13	13	2.00	Cytochrome P450 2C29 05=Mus musculus GN=Cyp2c29 PE=1
	2	SwissProt	#2::CP254_MOUSE	552	60887	27	27	8	8	0.88	Cytochrome P450 2C54 OS=Mus musculus GN=Cyp2c54 PE=2
	6	SwissProt	#2::CY250_MOUSE	489	61128	27	27	10	10	1.20	Cytochrome P450 2C50 OS=Mus musculus GN=Cyp2c50 PE=1
	4	SwissProt	2::CP2F2 MOUSE	484	59267	32	32	12	12	2.11	Cytochrome P450 2F2 OS=Mus musculus GN=Cyp2f2 PE=2 SV
	5	SwissProt	#2::CP237_MOUSE	339	60590	22	22	8	8	0.89	Cytochrome P450 2C37 OS=Mus musculus GN=Cyp2c37 PE=2
	6	SwissProt	#2::CP239_MOUSE	251	60856	13	13	4	4	0.37	Cytochrome P450 2C39 OS=Mus musculus GN=Cyp2c39 PE=2
	7	SwissProt	#2::CP238_MOUSE	150	61356	9	9	4	4	0.37	Cytochrome P450 2C38 OS=Mus musculus GN=Cyp2c38 PE=2
	1	SwissProt	#2::GRP78_MOUSE	1308	81404	55	55	21	21	2.47	78 kDa glucose-regulated protein OS=Mus musculus GN=Hspa
	2	SwissProt	#2::HSP7C_MOUSE	362	78937	21	21	8	8	0.63	Heat shock cognate 71 kDa protein OS=Mus musculus GN=Hs
	1	SwissProt	#2::CYBS_MOUSE	1217	16817	42	42	5	5	3.08	Cytochrome b5 OS=Mus musculus GN=Cyb5a PE=1 SV=2
	1	SwissProt	#2::PDIA1_MOUSE	1123	64694	53	53	16	16	2.54	Protein disulfide-isomerase OS=Mus musculus GN=P4hb PE=1
	1	SwissProt	#2::CP1A2_MOUSE	1054	63034	38	38	10	10	1.31	Cytochrome P450 1A2 OS=Mus musculus GN=Cyp1a2 PE=1 S
	1	SwissProt	#2::ENPL_MOUSE	1018	103744	63	63	10	19	1.53	Endoplasmin OS=Mus musculus GN=Hsp90b1 PE=1 SV=2
	1	SwissProt	#2::RDH7_MOUSE	1005	38455	45	45	12	12	4.07	Retinol dehydrogenase 7 OS=Mus musculus GN=Rdh7 PE=2 S
	2	SwissProt	#2::H1786_MOUSE	597	38949	23	23	7	7	1.37	17-beta-hydroxysteroid dehydrogenase type 6 OS=Mus muscu
	1	SwissProt	#2::MGST1_MOUSE	863	18595	25	25	3	3	2.57	Microsomal glutathione S-transferase 1 OS=Mus musculus GN
	1	SwissProt	#2::RL7A_MOUSE	770	35860	28	28	8	8	1.91	60S ribosomal protein L7a OS=Mus musculus GN=Rpl7a PE=2
	1	SwissProt	#2::RLA0_MOUSE	763	37215	24	24	7	7	1.47	60S acidic ribosomal protein P0 OS@Mus musculus GN@Rplp0
	1	SwissProt	#2::CP2AC_MOUSE	763	61325	35	35	14	14	2.25	Cytochrome P450 2A12 OS=Mus musculus GN=Cyp2a12 PE=1
	2	SwissProt	#2::CP2A5_HOUSE	59	61696	5	5	2	2	0.17	Cytochrome P450 2A5 OS=Mus musculus GN=Cyp2a5 PE=2 5
	1	SwissProt	#2::ACSL1_MOUSE	749	86078	38 15	38 15	18	18	1.90	Long-chain-fatty-acidCoA ligase 1 OS=Mus musculus GN=Ac
	2	SwissProt SwissProt	#2::ACSL5_MOUSE	297 748	84629 28083	31	15	6	7	0.41	Long-chain-fatty-acidCoA ligase 5 OS=Mus musculus GN=Ac
	1	SwissProt	#2::RL13_MOUSE #2::PDIA3_MOUSE	692	28083 64504	31 40	40	15	15	2.90	60S ribosomal protein L13 OS=Mus musculus GN=Rpl13 PE=2 Protein disulfide-isomerase A3 OS=Mus musculus GN=Pdia3 P
	1	SwissProt	#2::CP3AB_MOUSE	686	65154	32	32	10	10	1.25	Cytochrome P450 3A11 OS=Mus musculus GN=Cyp3a11 PE=1
	1	SwissProt	2::UDB17_MOUSE	677	67040	34	34	9	9	0.91	UDP-glucuronosyltransferase 2817 O5=Mus musculus GN=Ugt
	2	SwissProt	#2::UD11_MOUSE	429	05301	19	19	7	7	0.80	UDP-glucuronosyltransferase 1-1 OS=Mus musculus GN=Ugt1
	1	SwissProt	2::UD16_MOUSE	245	65516	14	14	6	6	0.67	UDP-glucuronosyltransferase 1-6 OS=Mus musculus GN=Ugt1
	li	SwissProt	2::EST3A_MOUSE	668	67490	28	28	5	5	0.43	Carboxylesterase 3A OS=Mus musculus GN=Ces3a PE=1 SV=2
	1.			1							

The Report Builder tab is useful when you need a table of proteins suitable for publication. Lets assume we want to drop the 'one hit wonders' and only report proteins that have significant matches to at least 2 different peptide sequences

				ed (3039)	2						<u>5 permalini</u>
	ı hi	ts (476 p	proteins)								
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ters	(00	one)									
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	Π.	Num of sign	ficant matches	1606	28266	48	48	7	7	2.86	sp[TRY1_BOVIN]
		Num of sequ	ences	1337	61419	76	76	13	13	2.00	Cytochrome P450 2C29 OS=Mus musculus GN=Cyp2c29 PE=
		Num of sign	licent sequences	552	60887	27	27	8	8	0.88	Cytochrome P450 2C54 OS=Mus musculus GN=Cyp2c54 PE=
		Num. of sign	ficant unique sequences	489	61128	27	27	10	10	1.20	Cytochrome P450 2C50 OS=Mus musculus GN=Cyp2c50 PE=
		emPAI Sequence co		484	59267	32	32	12	12	2.11	Cytochrome P450 2F2 OS=Mus musculus GN=Cyp2f2 PE=2 5
		pl	ireraya III	339	60590	22	22	8	8	0.89	Cytochrome P450 2C37 OS=Mus musculus GN=Cyp2c37 PE=
	l a	Description		251	60856	13	13	4	4	0.37	Cytochrome P450 2C39 OS=Mus musculus GN=Cyp2c39 PE= Cytochrome P450 2C38 OS=Mus musculus CN=Cyp2c39 PE=
		xed modifica Methylthia (C		150	61356	55	55	21	4	0.37	Cytochrome P450 2C38 OS=Mus musculus GN=Cyp2c38 PE= 78 kDa glucose-regulated protein OS=Mus musculus GN=Hsp
		ITRAQ4plex		362	78937	21	21	.8	8	0.63	Heat shock cognate 71 kDa protein OS=Mus musculus GN=H
	1	SwissProt	#2::CYBS_MOUSE	1217	16817	42	42	5	5	3.08	Cytochrome b5 OS=Mus musculus GN=Cyb5a PE=1 SV=2
	1	SwissProt	2::PDIA1_MOUSE	1123	64694	53	53	16	16	2.54	Protein disulfide-isomerase OS=Mus musculus GN=P4hb PE=
	1	SwissProt	#2::CP1A2_MOUSE	1054	63034	38	38	10	10	1.31	Cytochrome P450 1A2 OS=Mus musculus GN=Cyp1a2 PE=1
	1	SwissProt	#2::ENPL_MOUSE	1018	103744	63	63	19	19	1.53	Endoplasmin OS=Mus musculus GN=Hsp90b1 PE=1 SV=2
	1	SwissProt	#2::RDH7_MOUSE	1005	38455	45	45	12	12	4,07	Retinol dehydrogenase 7 OS=Mus musculus GN=Rdh7 PE=2
	2	SwissProt	#2::H1786_MOUSE	597	38949	23	23	7	7	1.37	17-beta-hydroxysteroid dehydrogenase type 6 OS=Mus musi
	1	SwissProt	#2::MGST1_MOUSE	863	18595	25	25	3	3	2.57	Microsomal glutathione S-transferase 1 OS=Mus musculus GI
	1	SwissProt	#2::RL7A_MOUSE	770	35860	28	28	8	8	1.91	60S ribosomal protein L7a OS=Mus musculus GN=RpI7a PE=
	1	SwissProt SwissProt	#2::RLA0_MOUSE	763	37215 61325	24 35	24 35	14	7	1.47	60S acidic ribosomal protein P0 OS=Mus musculus GN=Rplp0
	1	SwissProt	#2::CP2AC_MOUSE #2::CP2A5_MOUSE	59	61696	35	35	14	14	2.25	Cytochrome P450 2A12 OS=Mus musculus GN=Cyp2a12 PE= Cytochrome P450 2A5 OS=Mus musculus GN=Cyp2a5 PE=2
	1	SwissProt	#2::ACSL1_MOUSE	749	86078	38	38	18	18	1.90	Long-chain-fatty-acidCoA ligase 1 OS=Mus musculus GN=A
	2	SwissProt	#2::ACSL5_MOUSE	297	84629	15	15	6	6	0.41	Long-chain-fatty-acidCoA ligase 5 OS=Mus musculus GN=A
	1	SwissProt	#2::RL13_MOUSE	748	28083	31	31	7	7	2.90	605 ribosomal protein L13 OS=Mus musculus GN=Rpl13 PE=
	1	SwissProt	#2::PDIA3_MOUSE	692	64504	40	40	15	15	2.06	Protein disulfide-isomerase A3 OS=Mus musculus GN=Pdia3
	1	SwissProt	#2::CP3AB_MOUSE	686	65154	32	32	10	10	1.25	Cytochrome P450 3A11 OS=Mus musculus GN=Cyp3a11 PE=
	11	SwissProt	#2::UDB17 MOUSE	677	67040	34	34	9	9	0.91	UDP-glucuronosyltransferase 2B17 OS=Mus musculus GN=U

We open up the filters section and add a suitable filter.

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	1	CRAP	#1::spiTRY1_BOVINI	1606	28266	48	48	7	7	2.86	SPITRY1_BOVINI
	li	SwissProt	#2::CP2CT_MOUSE	1337	61419	76	76	13	13	2.00	Cytochrome P450 2C29 OS=Mus musculus GN=Cyp2c29 PE=1 5
	2	SwissProt	2::CP254 MOUSE	552	60887	27	27	8	8	0.68	Cytochrome P450 2C54 OS=Mus musculus GN=Cyp2c54 PE=2 Si
	5	SwissProt	#2::CY250_MOUSE	489	61128	27	27	10	10	1.20	Cytochrome P450 2C50 QS=Mus musculus GN=Cyp2c50 PE=1 S
	4	SwissProt	2::CP2F2 MOUSE	484	59267	32	32	12	12	2.11	Cytochrome P450 2F2 OS=Mus musculus GN=Cyp2f2 PE=2 SV=1
	5	SwissProt	#2::CP237_MOUSE	339	60590	22	22	8	8	0.89	Cytochrome P450 2C37 OS=Mus musculus GN=Cyp2c37 PE=2 S
	6	SwissProt	#211CP239_MOUSE	251	60856	13	13	4	4	0.37	Cytochrome P450 2C39 O5=Mus musculus GN=Cyp2c39 PE=2 5
	7	SwissProt	#2::CP238_MOUSE	150	61356	9	9	4	4	0.37	Cytochrome P450 2C38 OS=Mus musculus GN=Cyp2c38 PE=2 S
	1	SwissProt	#2::GRP78_MOUSE	1308	81404	55	55	21	21	2.47	78 kDa glucose-regulated protein OS=Mus musculus GN=Hspa5
	2	SwissProt	#2::HSP7C_MOUSE	362	78937	21	21	8	8	0.63	Heat shock cognate 71 kDa protein OS=Mus musculus GN=Hspal
	1	SwissProt	#2::CYB5_MOUSE	1217	16817	42	42	5	5	3.08	Cytochrome b5 OS=Mus musculus GN=Cyb5a PE=1 SV=2
	1	SwissProt	2::PDIA1_MOUSE	1123	64694	53	53	16	16	2.54	Protein disulfide-isomerase OS=Mus musculus GN=P4hb PE=1 SV
	1	SwissProt	#2::CP1A2_MOUSE	1054	63034	38	38	10	10	1.31	Cytochrome P450 1A2 OS=Mus musculus GN=Cyp1a2 PE=1 SV=
	1	SwissProt	#2::ENPL_MOUSE	1018	103744	63	63	19	19	1.53	Endoplasmin OS=Mus musculus GN=Hsp90b1 PE=1 5V=2
	1	SwissProt	21:RDH7_MOUSE	1005	38455	45	45	12	12	4.07	Retinol dehydrogenase 7 OS=Mus musculus GN=Rdh7 PE=2 SV=
	2	SwissProt	#2::H1786_MOUSE	597	38949	23	23	7	7	1.37	17-beta-hydroxysteroid dehydrogenase type 6 OS=Mus musculus
	1	SwissProt	#2::MGST1_MOUSE	863	18595	25	25	3	3	2.57	Microsomal glutathione S-transferase 1 OS=Mus musculus GN=M
1	1	SwissProt	2::RL7A_MOUSE	770	35860	28	28	8	8	1.91	60S ribosomal protein L7a OS=Mus musculus GN=Rpl7a PE=2 SV
( ) ( ) ( ) ( ) ( ) ( ) ( ) ( ) ( ) ( )	1	SwissProt	#2::RLA0_MOUSE	763	37215	24	24	7	7	1.47	60S acidic ribosomal protein P0 OS-Mus musculus GN-Rplp0 PE
	1	SwissProt	#2::CP2AC_MOUSE	763	61325	35	35	14	14	2.25	Cytochrome P450 2A12 OS=Mus musculus GN=Cyp2a12 PE=1 S
	2	SwissProt	#2::CP2A5_MOUSE	59	61696	5	5	2	2	0,17	Cytochrome P450 2A5 OS=Mus musculus GN=Cyp2a5 PE=2 SV=
	1	SwissProt	#2::ACSL1_MOUSE	749	86078	38	38	18	18	1.90	Long-chain-fatty-acidCoA ligase 1 OS=Mus musculus GN=AcsI1
	2	SwissProt	#21:ACSL5_MOUSE	297	84629	15	15	6	6	0.41	Long-chain-fatty-acidCoA ligase 5 OS=Mus musculus GN=AcsI5
	1	SwissProt	if2::RL13_MOUSE	748	28083	31	31	7	7	2.90	60S ribosomal protein L13 OS=Mus musculus GN=Rpl13 PE=2 S
	1	SwissProt	#2::PDIA3_MOUSE	692	64504	40	40	15	15	2.06	Protein disulfide-isomerase A3 OS=Mus musculus GN=Pdia3 PE=
	1	SwissProt	d2::CP3A8_HOUSE	686	65154	32	32	10	10	1.25	Cytochrome P450 3A11 OS=Mus musculus GN=Cyp3a11 PE=1 S
	1	SwissProt	#2::UDB17_MOUSE	677	67040	34	34	9	9	0.91	UDP-glucuronosyltransferase 2B17 OS=Mus musculus GN=Ugt2b
	2	SwissProt	#2::UD11_MOUSE	429	65361	19	19	7	7	0.80	UDP-glucuronosyltransferase 1-1 OS=Mus musculus GN=Ugt1a1
	3	SwissProt	#2::UD16_MOUSE	245	65516	14	14	6	6	0.67	UDP-glucuronosyltransferase 1-6 OS=Mus musculus GN=Ugt1a6
	1	SwissProt	#2::EST3A_MOUSE	668	67490	28	28	5	5	0,43	Carboxylesterase 3A OS=Mus musculus GN=Ces3a PE=1 SV=2

Only proteins with significant matches to at least 2 sequences remain. The filtering is very flexible, with lots of useful terms.

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	2	SwissProt	#2::CP254_MOUSE	552	60887	27	27	8	8	0.88	Cytochrome P450 2C54 OS=Mus musculus GN=Cyp2c54 PE=2 SV
	3	SwissProt		489	61128	27	27	10	10	1.20	Cytochrome P450 2C50 OS=Mus musculus GN=Cyp2c50 PE=1 SV
	4	SwissProt	#2::CP2F2_MOUSE	484	59267	32	32	12	12	2.11	Cytochrome P450 2F2 OS=Mus musculus GN=Cyp2f2 PE=2 SV=1
	5	SwissProt	#2::CP237_MOUSE	339	60590	22	22	8	8	0.89	Cytochrome P450 2C37 OS=Mus musculus GN=Cyp2c37 PE=2 SV
	6	SwissProt		251	60856	13	13	4	4	0.37	Cytochrome P450 2C39 OS=Mus musculus GN=Cyp2c39 PE=2 SV
	7	SwissProt	#2::CP238_MOUSE	150	61356	9	9	4	4	0.37	Cytochrome P450 2C38 OS=Mus musculus GN=Cyp2c38 PE=2 SV
	1	SwissProt	#2::GRP78_MOUSE	1308	81404	55	55	21	21	2.47	78 kDa glucose-regulated protein OS=Mus musculus GN=Hspa5 P
	-	SwissProt	2::HSP7C_MOUSE	362	78937			5		0.63	Heat shock cognate 71 kDa protein OS=Mus musculus GN=Hspa8
	1	SwissProt	#2::CYB5_MOUSE	1217	16817	42	42 53		5	3.08	Cytochrome b5 OS=Mus musculus GN=Cyb5a PE=1 SV=2
	1	SwissProt SwissProt	#2::PDIA1_MOUSE	1123	64694 63034	38	38	16	16	2.54	Protein disulfide-isomerase OS=Mus musculus GN=P4hb PE=1 SV
	1	SwissProt	2::CP1A2_MOUSE	1054	103744	63	63	10	10	1.31	Cytochrome P450 1A2 OS=Mus musculus GN=Cyp1a2 PE=1 SV=1 Endoplasmin OS=Mus musculus GN=Hsp90b1 PE=1 SV=2
	1	SwissProt	21:RDH7_MOUSE	1018	38455	45	45	19	19	4.07	Retinol dehydrogenase 7 OS=Mus musculus GN=Rdh7 PE=2 SV=2
	2	SwissProt	#2::H1786_MOUSE	597	38949	23	23	7	7	1.37	17-beta-hydroxysteroid dehydrogenase type 6 OS=Mus musculus
	1	SwissProt	#2::MGST1_MOUSE	863	18595	25	25	3	í í	2.57	Microsomal glutathione S-transferase 1 OS=Mus musculus GN=M
2	1	SwissProt	#2::RL7A_MOUSE	770	35860	28	28	8	8	1.91	60S ribosomal protein L7a OS=Mus musculus GN=Rpl7a PE=2 SV
	1	SwissProt	#2::RLA0_MOUSE	763	37215	24	24	7	7	1.47	605 acidic ribosomal protein P0 OS=Mus musculus GN=Rplp0 PE=
	1	SwissProt	2::CP2AC_MOUSE	763	61325	35	35	14	14	2.25	Cytochrome P450 2A12 OS=Mus musculus GN=Cyp2a12 PE=1 SV
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	2	SwissProt	#2::ACSL5_MOUSE	297	84629	15	15	6	6	0.41	Long-chain-fatty-acidCoA ligase 5 OS=Mus musculus GN=AcsI5
	1	SwissProt	#2::RL13_MOUSE	748	28083	31	31	7	7	2.90	60S ribosomal protein L13 OS=Mus musculus GN=Rpl13 PE=2 SV
	1	SwissProt	21:PDIA3_MOUSE	692	64504	40	40	15	15	2.06	Protein disulfide-isomerase A3 OS=Mus musculus GN=Pdia3 PE=1
	1	SwissProt	#2::CP3AB_MOUSE	686	65154	32	32	10	10	1.25	Cytochrome P450 3A11 OS=Mus musculus GN=Cyp3a11 PE=1 SV
	1	SwissProt	ef2::UD817_MOUSE	677	67040	34	34	9	9	0.91	UDP-glucuronosyltransferase 2817 OS=Mus musculus GN=Ugt2b1
	2	SwissProt	2::UD11_MOUSE	429	65361	19	19	7	7	0.80	UDP-glucuronosyltransferase 1-1 OS=Mus musculus GN=Ugt1a1 I
	3	SwissProt	#2::UD16_MOUSE	245	65516	14	14	6	6	0.67	UDP-glucuronosyltransferase 1-6 OS=Mus musculus GN=Ugt1a6 R
	1	SwissProt	#2::EST3A_MOUSE	668	67490	28	28	5	5	0.43	Carboxylesterase 3A OS=Mus musculus GN=Ces3a PE=1 SV=2
	1	SwissProt	#2::RL4_MOUSE	650	55568	34	34	11	11	1.59	605 ribosomal protein L4 OS=Mus musculus GN=Rpl4 PE=1 SV=3

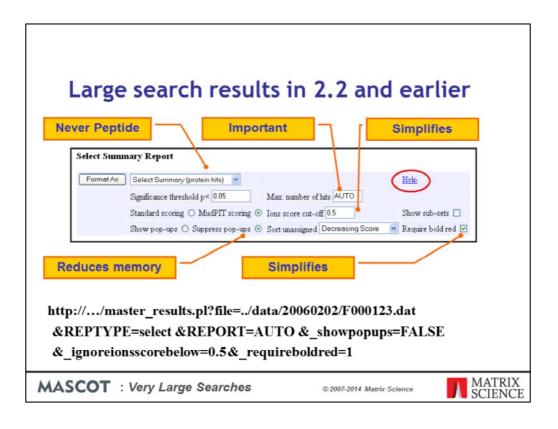
Another thing that you could easily do would be to exclude proteins from the contaminants database

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		12 out of 16)									
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2	SwissProt			552	60887	27	27	8	8	0.88	Cytochrome P450 2C54 OS=Mus musculus GN=Cyp2c54 PE=2 5
3	SwissProt			489	61128	27	27	10	10	1.20	Cytochrome P450 2C50 OS=Mus musculus GN=Cyp2c50 PE=1 :
4 5	SwissProt SwissProt			484	59267 60590	32 22	32	12	12	2.11 0.89	Cytochrome P450 2F2 OS=Mus musculus GN=Cyp2f2 PE=2 SV= Cytochrome P450 2C37 OS=Mus musculus GN=Cyp2c37 PE=2 9
6	SwissProt			251	60856	13	13	4	4	0.89	Cytochrome P450 2C37 OS=Mus musculus GN=Cyp2c37 PE=2 Cytochrome P450 2C39 OS=Mus musculus GN=Cyp2c39 PE=2
7	SwissProt			150	61356	9	13	4	4	0.37	Cytochrome P450 2C39 OS=Mus musculus GN=Cyp2c39 PE=2 5 Cytochrome P450 2C38 OS=Mus musculus GN=Cyp2c38 PE=2 5
1	SwissProt			1308	81404	55	55	21	21	2.47	78 kDa glucose-regulated protein OS=Mus musculus GN=Hspa5
2	SwissProt	2::HSP7C		362	78937	21	21	8	8	0.63	Heat shock cognate 71 kDa protein OS=Mus musculus GN=Hsoa
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The columns section of Report Manager allows you to choose which columns to include and, if required, change their order

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30							matches	significant matches	sequences	significant		
31		1	iPRG_2012	P00925	2140	46942	148	100	53	43	44.71	Enclase 2 OS=Saccharomyces cere
32		2	iPRG_2012	P00924	1059	46844	71	46	35	27	7.47	Enolase 1 OS=Saccharomyces cere
33		1	iPRG_2012	P00549	1933	54909	133	87	56	43	18.28	Pyruvate kinase 1 OS=Saccharomyc
34		1	PRG_2012	P40150	1613	66668	105	66	66	45	11.76	Heat shock protein SSB2 OS=Sacch
35 36		2	PRG_2012	P11484 P10592	1590 1591	66732 69599	103	65 57	64 52	44 32	11.12	Heat shock protein SSB1 OS=Sacch Heat shock protein SSA2 OS=Sacch
30 37		2	iPRG 2012	P10592	1161	69786	85	44	48	26	3.02	Heat shock protein SSA2 US=Sacch Heat shock protein SSA1 OS=Sacch
38		3	PRG 2012	P16474	233	74479	23	8	17	6	0.32	78 kDa glucose-regulated protein hor
39		1	IPRG 2012	P00330	1453	37282	73	51	32	25	13.48	Alcohol dehydrogenase 1 OS=Sacch
40		2	IPRG_2012	P07245	101	40743	14	5	7	3	0.29	Alcohol dehydrogenase 3, mitochono
41		1	PRG 2012	P00560	1382	44768	102	58	54	33	12.75	Phosphoglycerate kinase OS=Sacch
42		1.	IPRG 2012	P00359	1361	35838	76	54	31	25	12.29	Glyceraldehyde-3-phosphate dehydro
43	7	2	PRG 2012	P00358	1242	35938	69	48	29	24	9.89	Glyceraldehyde-3-phosphate dehydro
44	7	3	iPRG_2012	P00360	505	35842	30	20	14	12	2.47	Glyceraldehyde-3-phosphate dehydro
45		4	iPRG_2012	P04406	41	36201		2	4	2	0.21	Glyceraldehyde-3-phosphate dehydro
46		1	(PRG_2012	P06169	1289	61685		41	28	26	4.7	Pyruvate decarboxylase isozyme 1 C
47		1	iPRG_2012	P00950	1031	27592		44	32	25	34.97	Phosphoglycerate mutase 1 OS=Sar
48		1	iPRG_2012	P07281	1015	15881	51	38	16	13	22.71	40S ribosomal protein S19-B OS=Sa
49		2	PRG_2012	P07280 P00761	1014 922	15907 25078	51 37	38 27	16 7	13 6	22.71 2.89	40S ribosomal protein S19-A OS=Sa
50 51		1	iPRG 2012	P32324	784	25070		33	33	23	1.44	SWISS-PROT:P00761[TRYP_PIG Tr Elongation factor 2 OS=Saccharomy
52		1	PRG 2012	P16521	771	116727		33	47	30	1.52	Elongation factor 2 US=Saccharomy Elongation factor 3A US=Saccharomy
53		1	IPRG 2012	P05319	765	10739	38	29	10	9	95.65	60S acidic ribosomal protein P2-alph
54		1	iPRG 2012	003048	721	15948	28	23	17	14	17.82	Cofilin OS=Saccharomyces cerevisia
55		1	IPRG_2012	P0C0V8	719	9797	42	29	15	12	207.43	40S ribosomal protein S21-A OS=Sa
56	16	2	PRG_2012	03E754	694	9811	41	28	15	12	148.28	40S ribosomal protein S21-B OS=Sa
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Once the list is filtered and the columns arranged as required, there is a button to export the table as CSV, which can be pasted into Excel and formatted to create a suitable figure for dropping into a publication



If you are still using Mascot 2.2 or if you have some application software that requires the results in the earlier format, and you are encountering problems with timeouts and running out of memory, here are some tips:

•Ensure you are using the Select report. If you are using a third party client that has specified Peptide summary or Protein summary, add this to the URL before opening the file: &REPTYPE=select

•Don't specify a huge number of hits 'just in case'. Choose AUTO to display all protein hits that contain at least one significant peptide match: &REPORT=AUTO

•Get rid of the yellow pop-ups: &\_showpopups=FALSE

•Setting require bold red and an expect value cut-off will minimise the number of hits: &\_ignoreionsscorebelow=0.5&\_requireboldred=1

Note that the ions score cut-off is as score threshold when the value is 1 or greater. When the value is between 0 and 1, it is an expect threshold, which is often much more useful. I often set this to 0.5 to get rid of all the junk matches.

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master_results.pl					
URL	mascot.dat	Value	Description		
		peptide	Peptide Summary		
		archive	Archive Report		
reptype		concise	Concise Protein Summary		
		protein	Full Protein Summary		
		select	Select Summary (hits)		
		unassigned	Select Summary (unassigned)		
report		auto	Report all significant hits		
i opore		N	Report N hits		
_showsubsets	ShowSubSets	1	Set value to 1 to report Peptide Summary hits that match a subset of peptides. Default is 0.		
_requireboldred	RequireBoldRed	1	Set value to 1 to report Peptide Summary hits only if they contain at least one "bold red" peptide. Default is 0.		
_showallfromerrortolerant	ShowAllFromErrorTolerant	1	Set value to 1 to report all hits from an error tolerant search, including the garbage. Default is 0.		
_sigthreshold	SigThreshold	N	Probability to use for the significance threshold. Range is 0.1 to 1E-18. Default is 0.05.		
		scoredown	Sort unassigned matches by descending score, (default)		
_sortunassigned	SortUnassigned	queryup	Sort unassigned matches by ascending query number		
		intdown	Sort unassigned matches by descending intensity		
_ignoreionsscorebelow	IgnoreIonsScoreBelow	N	Any ions scores below this value are set to 0. Floating point number, default 0.0.		
_showpopups		true	Show top 10 peptide matches fro each query in JavaScript pop-up, (default)		
		false	Suppress JavaScript pop-ups.		
_alwaysgettitle		1	Set to 1 to force reports to fetch Fasta titles from database when they are not included in the result file. Default is 0.		
_mudpit	Mudpit	N	Number of queries at which protein score calculation switches to large search mode. Default 1000		

If you can't remember these URL parameters, just click on the help link

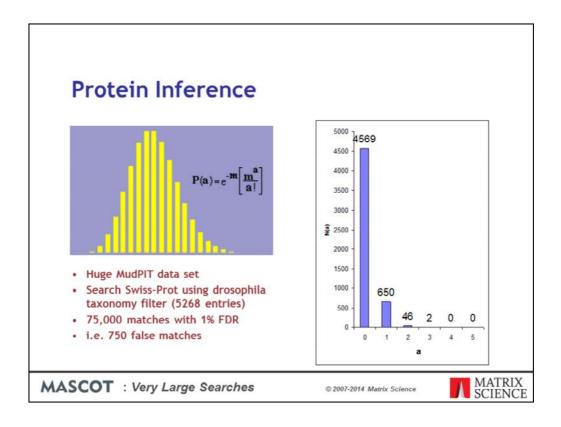
Select Summary Report         Format As       Select Summary (protein hits)         Standard scoring       Max number of hits AUTO         Standard scoring       MudPIT scoring       Ions score cut-off         Show pop-ups       Suppress pop-ups       Sort unassigned	Help Show sub-sets Require bold red
MASCOT : Very Large Searches © 2007-2014 Matrix Scient	MATRIX SCIENCE

What do we mean by Standard scoring and MudPIT scoring?

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		Observed	Mr(expt)	Mr(calc)			Score	Expect	Rank	Unique	Peptide	
	28	359.7341	717.4537	717.4537	-0.09	0	7	4.2	5	U	R.LFAIVR.G	
		394.2371	786.4596			0	8	13	3	U	K.LTIADVR.A	
		411.2073	820.4000	820.3954	5.61	0	3	15	4	U	K. TDSGLYR.C	
							12	1.1	5	U	K. RFLTLR. K	
	357	413.2642	824.5139		0.48	1						
	357 715	413.2642 450.7365	899.4584	899.4588	-0.38	0	10	2.9	2	U	K. IVDVSSDR.C	
	357 715 740	413.2642 450.7365 451.7681	899.4584 901.5217	899.4588 901.5233	-0.38	0	10 3	2.9	2 3	U	K. IVDVSSDR.C R.VTLVDVTR.N	
	357 715 740 840	413.2642 450.7365 <b>451.7681</b> 459.2484	899.4584 901.5217 916.4821	899.4588 901.5233 916.4767	-0.38 -1.72 5.98	0	10 3 2	2.9 24 29	2 3 2	U U	K. IVDVSSDR.C R.VTLVDVTR.N R.GVEFNVPR.L	
	357 715 740 840 844	413.2642 450.7365 <b>451.7681</b> 459.2484 459.7299	899.4584 901.5217 916.4821 917.4452	899.4588 901.5233 916.4767 917.4454	-0.38 -1.72 5.98 -0.24	0 0 0	10 3 2 4	2.9 24 29 15	2 3 2 6	<b>บ</b> บ บ	K. IVDVSSDR.C R.VILVDVIR.N R.GVEFNVPR.L K.ELEETAAR.N	
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	357 715 740 840 844 1029 1056	413.2642 450.7365 451.7681 459.2484 459.7299 473.2757	899.4584 901.5217 916.4821 917.4452 944.5368	899.4588 901.5233 916.4767 917.4454 944.5331 949.4869	-0.38 -1.72 5.98 -0.24 3.97	0 0 0 1	10 3 2 4 3	2.9 24 29 15 21	2 3 2 6 3	ม ช ม	K. IVDVSSDR.C R.VILVDVIR.N K.GVEFNVPR.L K.ELEETAAR.M R.EPPSFIKK.I	

With standard peptide summary scoring, the protein score is essentially the sum of the ions scores of all the peptides assigned to the protein. Where there are duplicate matches to the same peptide, the highest scoring match is used. A correction is applied based on the number of candidate peptides that were tested. This correction is very small unless it is a very large protein, like here, or a no-enzyme search

Despite this correction, as this example shows, when we have many low scoring matches assigned to the same protein, we can still get a high protein score, even though none of the individual peptide matches are significant



A protein with matches to just a single peptide sequence is commonly referred to as a "one-hit wonder" and is often treated as suspect. This is actually a slight oversimplification. In a search with a large number of spectra and a small database, even though the peptide false discovery rate is low, a protein can pick up multiple false matches by chance. This is easily calculated using a Poisson Distribution, where m is the average number of false matches per protein. In this example, m is 750/5268, and we would expect 650 database entries to be one-hit wonders. However, 46 entries will pick up two false matches and 2 entries will pick up three, which could mean we report 48 false proteins.

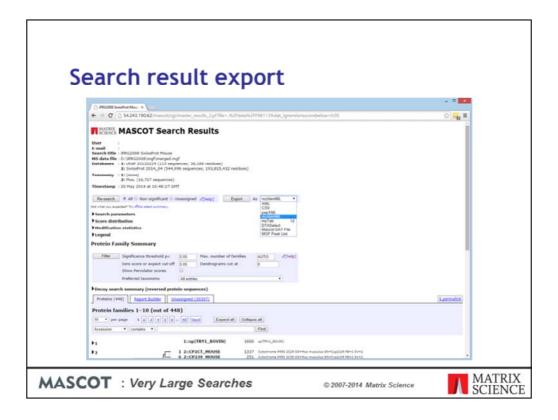
The problem isn't limited to large searches. It is the ratio between the number of spectra and the number of entries in the database that matters. So, a small search against a small database can give similar numbers

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28	359.7341 394.2371		717.4537	-0.09	0	7	4.2	5	U	R.LFAIVR.G
209	394.2371 411.2073	786.4596		-0.46	0	8	13	4	0	K.LTIADVR.A
334	411.2073	820.4000		0.48	1	12		4	U	K.TDSGLYR.C K.RFLTLR.K
	413.2642	824.5139		-0.38	0	10		2	U	K. IVDVSSDR.C
	451.7681	901.5217		-1.72		3		3	u	R.VILVDVIR.N
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740 840 844 1029 1058	459.2484 459.7299 <b>473.2757</b> 475.7505	917.4452 944.5368 949.4864	916.4767 917.4454 944.5331 949.4869	-0.24 3.97 -0.47	0 1	4 3 4	15 21 22	3	U	K.ELEETAAR.M R.EPPSFIKK.I R.SSVSLSVCK.P
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740 840 844 1029 1058	459.2484 459.7299 473.2757 475.7505 476.2790	917.4452 944.5368 949.4864 950.5433	916.4767 917.4454 944.5331 949.4869 950.5425	-0.24 3.97 -0.47 0.94	0 1	4 3 4	15 21 22	3 5	U U	K.ELEETAAR.M R.EPPSFIKK.I R.SSVSLSVCK.P

To avoid this problem, we use MudPIT protein scoring, in which the score for each peptide match is not its absolute score, but the amount that it is above the threshold. Therefore, matches with a score below the threshold do not contribute to the score. The MudPIT protein score is the sum of the score excess over threshold for each of the matching peptides plus one times the average threshold. For each peptide, the "threshold" is the homology threshold if it exists, otherwise it is the identity threshold.

So, even though a large protein like titin may pick up several random matches, with MudPIT scoring, the protein score is zero, so you don't see it listed in the report unless you specify a huge number of protein hits, as was done here to capture this screen shot.

By default, MudPIT protein scoring is used when the ratio between the number of queries and the number of database entries, (after any taxonomy filter), exceeds 0.001. This default switching point can be moved by changing the value of MudpitSwitch in mascot.dat. You can also switch between the two scoring methods by using the format controls at the top of the report.



At some stage, it is likely that you will want to export the search results to another application or a relational database. If you want to write your own code, we provide a free library called Mascot Parser that provides a clean, object oriented programming interface to the result file. The supported languages are C++, Java, and Perl.

Mascot also includes a flexible export utility.

If you want the XML format, you probably know that this is what you want. If you've no idea what XML is, chances are you don't want it.

Choose CSV if you want to export to Excel - I'll show an example in a moment.

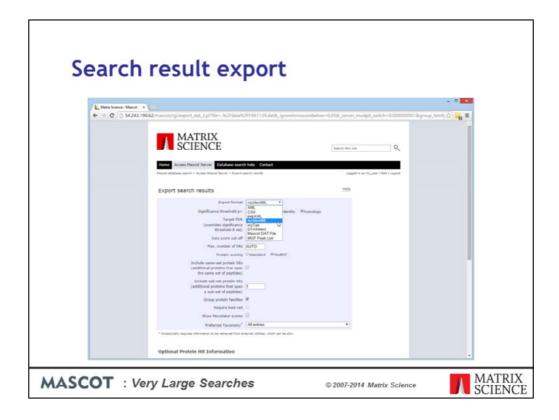
Choose pepXML if you want to export to Protein Prophet from ISB.

mzIdentML and mzTab are the standard formats from PSI for search result interchange. Mascot provides a very full implementation of mzIdentML and this is the one to choose if you are writing new application software that will use Mascot results

DTASelect is the tab separated format used by David Tabb's DTASelect program

The Mascot DAT file is the raw result file. If you need the result file for some reason, and don't have FTP or SCP access to your Mascot server, this is a convenient way to get the file.

MGF peak list is useful when you have the search result but can't find the peak list.



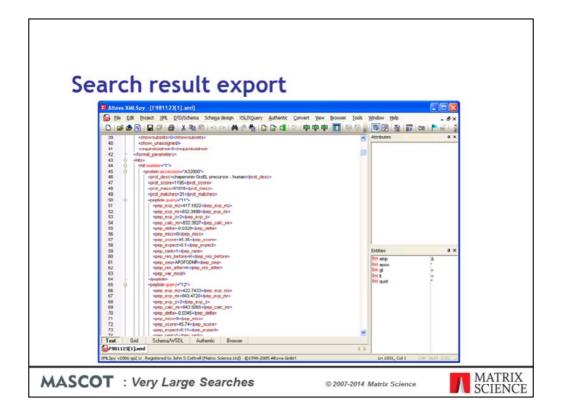
If you arrive here from one of the older reports, to begin with, you may need to select the required output format. Different formats have different options further down the page



To export to Excel, simply select CSV as the format, and click on the Export Search Results button at the bottom of the page. In recent versions of Mascot, the report is prepared and then a download button is displayed. In older versions, the download would start immediately. One the download is finished, you can open it into Excel:

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49 1					1000	12	422.7433	843.472	2	843.5065	-0.0345	
50 1							430.7328			859.4837	-0.0327	
51 1		_					451.2499		2		-0.0427	
52 1 53 1							456.7806 480.7447			911.5803 959.5036		
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Much easier and safer than "screen scraping"



For those of you into XML, here is a sample XML file. The schema is available from our web site or your local Mascot installation.

Please read the help for details.

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456.7806	911.5467	2	911.5803	-0.0337	Ő	59	0.0041	1	ĸ	VGLQVVAVK	A
490.7447	959.4748	2	969.5036	-0.0288	0	45.33	0.11	14	R	VTDALNATR	A
595.7855	1189.5565	2	1189.6012	-0.0447	0	56.55	0.0068	1	K	EIGNISDAMK	ĸ
603.7720	1205.5294	2	1205.5961	-0.0668	0	50.13	0.027	1	K	EIGNIISDAMK	к
608.3099	1214.6052	2	1214.6506	0.0454	0	73.21	0.00015	1	K	NAGVEGSLIVEK	1
617 2057	1232.5569	2	1232.5884	-0.0315	0	80.63	2.7e-05	1	ĸ	VOGTSDVEVNEK	к
672.8375	1343.6605	2	1343.7085	-0.0480	0	64.38	0.001	1	R	TVIEQSWGSPK	V _
714.0084	1427.7623	2	1427.8057	-0.0434	0	64.52	0.00086	1	R	GVMLAVDAVIAELK	к
714.8938	1427.7730	2	1427.8057	-0.0327	0	72.61	0.00013	1	R	GVMLAVDAVIAELK	K
722.8849	1443.7552	2	1443.8006	-0.0454	0	72.71 70.08	0.00014	1	R	GVMLAVDAVIAELK GVMLAVDAVIAELK	ĸ
752.8643	1443.7722	2	1503,7490	-0.0264	0	10.06	2.7e-06	6	K	TLNDELEIEGMK	F
760.8461	1519.6777	2	1519,7439	-0.0662	0	84.43	8.9e-06	1	K	TLNDELEIEGMK	F
640.3281	1917.9625	3	1918.0636	-0.1010	0	101.5	1.3e-07	1	K	ISSIGSTVPALEIANAHR	ĸ
960.0327	1918.0509	2	1918.0636	-0.0127	0	87.34	3.2e-06	1	к	ISSIQSIVPALEIANAHR	K
1019.5106	2037.0067	2	2037.0153	-0.0086	0	52.42	0.01	1	R	IGEIEGLOVITSEYEK	E
1057.0537	2112.0929	2	2112.1322	-0.0393	0	115.78	4.6e-09	1	R	ALMLQGVDLLADAVAVTMGPK	
1065.0399	2128.0653	2	2128.1271	-0.0618	0	68.73	0.00022	1	R	ALMLOGVDLLADAVAVTMGPK	
1073.0477 789.1062	2144.0809 2364.2968	2	2144.1220 2364.3263	-0.0411	0	69.64 55.53	0.00018	1	R	ALMLQGVDLLADAVAVTMGPK KPLVIAEDVDGEALSTLVLNR	G
1183.1570	2364 2966	2	2364.3263	-0.0296	0	65.46	0.00038	1	R	KPLVIAEDVDGEALSTLVLNR	1
789.1094	2364 3063	3	2364 3263	-0.0200	ő	94 59	4.5e-07	1	R	KPLVIAEDVDGEALSTLVLNR	i.
030 1277	7401 2740	2	14.00 104.0	0.0103	n	17 63	0.072		0	TALLDA AGUACH TTAELAA/TE	
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XML is ideal for transferring the results to a relational database. Even Microsoft Access can open the XML file directly into database tables

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6 PRINTING		Jan Ville							* Ites al description. *

There is a very detailed help page for all of this.

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Which describes how the export script can be called from the command line or a shell prompt, as part of an automated pipeline.

I won't go into any detail here, but this means that it is possible to set up a script that will, for example, automatically convert all of your Mascot results to XML files.

Figuring out the command line arguments from the help can be tricky so, in Mascot 2.3, we added a function to display the command line corresponding to the selected options



By the way, don't delete the original result files after exporting them or your won't be able to view the standard Mascot reports in a browser.