

MASCOT

MATRIX SCIENCE



Very large searches present a number of challenges. These are the topics we will cover during this presentation.

The smartest way to merge files, like fractions from a fractionated run, is using Mascot Daemon. Just tick the box at the bottom left.

The batch can be peak lists or raw files

Note that Mascot Daemon 2.1 had a file size limit of 2 GB. This was lifted in 2.2, and we have successfully merged and searched a 6 GB file on a Linux system.

For windows web servers the upload limit is 4 GB. In Mascot Daemon 2.6 we introduced an option to run searches from the command line if Mascot Daemon and Mascot Server are installed on the same computer. This bypasses any web server file limit and search sizes are effectively unlimited.

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Now Mascot Daemon 2.7 gives you another way to merge searches.

Instead you can select multiple searches in a Mascot Daemon task by CTRL+click individually searches or shift+click a range then right click and choose combined report.

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The combined search will open in a web page and list the results files that have been merged at the top of the report.

This will work with searches that have been processed by any peak picking software including Mascot Distiller.

Data	files		
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	Concatenating peal	k lists:	
	• DTA or PKL		
	Download merge.pl front http://www.matrixscie	om the Matrix Science Xcalibur help page nce.com/help/instruments_xcalibur.html	
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If you don't want to use Daemon, you can merge peak lists manually.

For DTA or PKL, you can download a script from our web site.

A nice feature of this script is that it puts the filename into the scan title, so you can tell which fraction a particular spectrum came from. The scan titles are displayed in the yellow pop-ups on the Mascot result report

Data files		
•MGF		
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As long as MGF files contain only peak lists, you don't need a script. Just use copy or cat

If the MGF files have search parameters at the beginning, you'll need to remove these before merging the files. Because a number of third party utilities add commands to MGF headers, and these cause a merged search to fail, Mascot Daemon 2.3 and later strips out header lines when merging MGF files.



In talking to Mascot users, it is clear that peak lists files are often much bigger than they should be. In other words, the peak detection is not very good. If you do a back of the envelope calculation, you can see that 2 GB should be enough for approximately 1 million spectra.

If you intend to do a lot of large searches, its worth getting the peak detection right. Shipping unnecessarily large files around wastes both time and disk space



32 bit platforms have a maximum process size of 2 GB on Windows or 3Gb on Linux. To get around this limit, Mascot divides large searches into smaller chunks, so as to avoid having everything in memory at the same time. The parameters to control this are SplitNumberOfQueries and SplitDataFileSize in the Options section of mascot.dat

One consequence of splitting a search is that there is no protein summary section in the result file. This is not a problem, because no-one wants a protein summary report for a large MS/MS search. However, some old client software gets confused by the missing section. The work around is to increase the values so that large searches never split. Maybe setting SplitNumberOfQueries to 1 million spectra and SplitDataFileSize to 10 billion bytes.

This is OK, but remember to reset these values as soon as you are able to. Otherwise, you might find you run out of memory or address space for your large searches

Mascot Server is now fully 64 bit. If you have enough RAM you could avoid splitting the search into chunks, however we still do by default because there is no performance penalty in doing so.



In early versions of Mascot, trying to display result reports for very large searches would often lead to problems with timeouts and running out of memory. To address this, the Protein Family Summary loads most of the information 'on demand'. This requires some index files to be created on the server, and these index files are cached, so that the report loads much faster on the second and subsequent occasions. Proteins are grouped into families by means of shared peptide matches and, within each family, hierarchical clustering is used to illustrate which proteins are closely related and which are more distant.

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	3 2::CY250_MOUSE	490	Cytochrome P450 2C50 OS=Mus musculus GN=Cyp2c50 PE=1 SV=2	
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If there are 300 or more spectra, the Protein Family Summary is the default. This is the appearance of a typical family report immediately after loading. The body of the report consists of three tabs, one for protein families, one for Report Builder, and one for unassigned matches. The report is paged, with a default page size of 10 families. If you wish, you can choose to display a larger number of families on a single page.

Proteins are grouped into families using a novel hierarchical clustering algorithm. If the family contains a single member, the accession string, protein score and description are listed. If the family contains multiple members, the accessions, scores and descriptions are aligned with a dendrogram, which illustrates the degree of similarity between members.

The scores for the proteins in family 2 vary from 1337 down to 73. In the earlier Peptide Summary or Select Summary reports, these would have been at opposite ends of the report. It would have been difficult to recognise that these proteins belonged together, even though they have shared peptide matches and are all cytochrome P450.

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If you are interested in family 2, then you click to expand it to show the details.

Immediately under the dendrogram is a list of the proteins. The table of peptide matches is similar to that found in the other result reports. We only reports the significant peptide matches less than the p value for the search that defaults to p<0.05. Duplicate matches to the same sequence are collapsed into a single row. The columns headed 1, 2, 3, etc. represent the proteins and contain a black square if the peptide is found in the protein. Some matches are shared, but each protein has some unique peptide matches, otherwise it would be dropped as a sub-set.

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Moving down to family 3, the scale on the dendrogram is ions score, and HSP7C_MOUSE and HS71L_MOUSE join at a score of approximately 30. This represents the score of the significant matches that would have to be discarded in order to make one protein a sub-set of the other. These two proteins are much more similar to one other than to GRP78_MOUSE, which has non-shared peptide matches with a total score of approximately 145. Note that, where there are multiple matches to the same peptide sequence, (ignoring charge state and modification state), it is the highest score for each sequence that is used.

Immediately under the dendrogram is a list of the proteins. In this example, because SwissProt has low redundancy, each family member is a single protein. In other cases, a family member will represent multiple same-set proteins. One of the proteins is chosen as the anchor protein, to be listed first, and the other same-set proteins are collapsed under a same-set heading. There is nothing special about the protein picked for the anchor position. You may have a preference for one according to taxonomy or description, but all proteins in a same-set group are indistinguishable on the basis of the peptide match evidence.

The table of peptide matches is similar to that found in the other result reports. Duplicate matches to the same sequence are collapsed into a single row. Click on the triangle to expand.

The black squares to the right show which peptides are found in which protein. To see the peptides that distinguish HSP7C_MOUSE and HS71L_MOUSE, clear the checkbox for GRP78 MOUSE and choose Redisplay.

▼3 1 2::GRP78_MOUSE 2 2::HSP7C_MOUSE 3 2::HSP7L_MOUSE 1308 78 lba glucese-regulated protein 05=Mus musculus 0N=Hspas PE=1 SV=1 362 362 Heat block regulated protein 05=Mus musculus 0N=Hspas PE=1 SV=1 188 362 Heat block rog notein 1-like 05=Mus musculus 0N=Hspas PE=2 SV=4 4 > > > > > > >	
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▶5 2::PDIA1_MOUSE 1123 Protein disuffde-isomerase OS=Hus musculus GN=PAbb PE=1 SV=2	
>6 2::CP1A2_MOUSE 1054 Crysterheame R450 1A2 05=Mus musculus 0N=Cyp2a2 REv15V=1	
▶7 2::ENPL_MOUSE 1018 Endoplasmin OS=Mus musculus GN=Hap9061 PE=1 SV=2	-

It can now be seen that HS71L_MOUSE would be a sub-set of HSP7C_MOUSE if it was not for one match, K.ATAGDTHLGGEDFDNR.L. It is the significant score for this match that separates the two proteins in the dendrogram by a distance of 32 (score of 55 - homology threshold score of 23).

You can "cut" the dendrogram using the slider control.

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#5924) 1	546.9979 1091.9813 1091.6430	0.3383 0 41 0	.0061)1 K.ITITNDK.G	
zf 6994	565.8689 1129.7232 1129.6101	0.1131 0 32	0.027 🕨 🔳 U 🔳 R.LTPEEIER.M	
m 7519	573.9761 1145.9377 1145.6536	0.2841 0 38 0	.0061 1 U R.GTLDPVEK.A	
# 9021	596.5726 1191.1306 1190.6725	0.4581 0 45 0	.0028 1 U R.VMEHFIK.L	
₫9459 ▶2	603.8705 1205.7264 1205.6747	0.0517 0 61 5.		
2 2557 13	609.9429 1217.8713 1217.6486	0.2227 0 45 0	0001 K. H. K. KUNDITEL	
10037 11	612,8115 1835,4126 1834,8204	0.5923.0 35 0	.0075 U K.STAGDTHLOGRDFDNR.M	
#11545	635.4900 1268.9654 1268.6856	0.2799 0 55 0.1	00058 U K.ETAEAYLGK.K	
sf11946 þ1	641.5476 1281.0806 1280.7220	0.3586 0 55 0.0	00015 1 U K.EIAEAYLGK.T	
ef18194	740.5968 1479.1791 1478.8336	0.3455 1 42 0	.0011 1 U K.VYEGERPLTK.D	
ef18197	494.1957 1479.5652 1478.8336	0.7316 1 26	0.014 1 U K.VYEGERPLTK.D	
z*19656	769.0064 2303.9974 2303.2630	0.7344 1 39 0	.0053 🕨 U 🔳 K.KVTHAVVTVPAYFNDAQR.Q	
m21354 1	803.1397 1604.2648 1603.8337	0.4311 0 63 3.3	2e-05 1 U R.NELESYAYSLK.N	
ef22754	558.5383 1672.5930 1671.9231	0.6699 1 28	0.018 1 U K.MKETAEAYLOK.K	
±23465 ▶2	855.9392 1709.8639 1709.8746	-0.0108 0 66 2.3	1e-06 1 U R.ITPSYVAFTPEGER.L	
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If we cut the dendrogram at a score of 50, HS71L_MOUSE will be dropped because it is now a sub-set protein. If you compare the matches to HSP7C_MOUSE with those to GRP78_MOUSE, it is clear that these are very different proteins. They are part of the same family because of two shared matches, but many highly significant matches would have to be discarded for either protein to become a sub-set of the other. In summary, we can quickly deduce from the Family Summary that there is abundant evidence that both GRP78_MOUSE and HSP7C_MOUSE were present in the sample. There is little evidence for HS71L_MOUSE. It is more likely that the HSP7C_MOUSE contained a SNP or two relative to the database sequence.

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41 2::NB5R3_MOUSE	364 NADH-cytochrome b5 reductase 3 OS=Mus musculus GN=Cyb5r3 PE=1 SV=3	
42 2::RS19_MOUSE	360 40S ribosomal protein S19 OS=Mus musculus GN=Rps19 PE=1 SV=3	
▶43 2::CP2E1_MOUSE	358 Cytochrome P450 2E1 OS=Mus musculus GN=Cyp2e1 PE=2 SV=1	
▶44 2::RL22_MOUSE	347 60S ribosomal protein L22 OS=Mus musculus GN=Rpl22 PE=1 SV=2	
▼45 2::RS15A_MOUSE	344 405 ribosomal protein 515a OS=Mus musculus GN=Rps15a PE=1 SV=2	
45.1 #2::RS15A_MOUSE	Score Mass Matches Sequences emPAI 344 16651 16 (16) 3 (3) 2.12 405 ribosomal protein S15a OS=Mus musculus GN=Rps15a PE=1 SV=2	
▼16 peptide matches (4 non-duplicate,	12 duplicate)	
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211283 631.9416 126	51.8686 1261.7308 0.1379 0 (59) 0.00013 1 U R. MNVLADALK.S	
632.0080 126	52.0014 1261.7308 0.2706 0 (42) 0.0065 1 U.R. MNVLADALK.S	
z11604 1 636.4751 127	10.9355 1270.6904 0.2452 0 28 0.03 1 U K.WQNNLLPSR.Q	
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£11790 639.9899 127	77.9652 1277.7257 0.2396 0 (48) 0.00054 ▶1 U R.MMVLADALK.S + Oxidatica (M)	
	223 URA-burnersburgforer 113 Afeller marche Ateliatist Met Atel	
47 2::002A3_MOUSE	3.3.3 vvv-gvvvrovosynamiser38 2.4.3 US=Nus musculus On=Vugt23 PE=1 SV=1	
48 2::FM05_MOUSE	315 Dimethylaniline monooxygenase [N-oxide-forming] 3 QS=Mus musculus GN=Fmo5 PE=2 SV=4	
	314 40S ribosomal protein S9 OS=Mus musculus GN=Rps9 PE=1 SV=3	
▶ 49 2::RS9_MOUSE		

The family report also includes a text search facility, which is particularly important for a paged report. You can search by accession or description sub-string, or by query, mass or sequence. Here, for example, we searched for a peptide sequence. The display jumps to the first instance of the sequence, expands, and highlights (in green) the target peptides.

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2	2	SwissProt	12::CP254 MOUSE	552	60887	27	27	8	8	0.88	Cytochrome P450 2C54 OS=Mus musculus GN=Cyp2c54 R	E=2 :
2	3	SwissProt	#2::CY250 MOUSE	489	61128	27	27	10	10	1.20	Cytochrome P450 2C50 OS=Mus musculus GN=Cyp2c50 B	E=1:
2	4	SwissProt	2::CP2F2 MOUSE	484	59267	32	32	12	12	2.11	Cytochrome P450 2F2 OS=Mus musculus GN=Cyp2f2 PE=	2 SV:
2	5	SwissProt	#2::CP237_MOUSE	339	60590	22	22	8	8	0.89	Cytochrome P450 2C37 OS=Mus musculus GN=Cyp2c37 F	E=2 :
2	6	SwissProt	#2::CP239_MOUSE	251	60856	13	13	4	4	0.37	Cytochrome P450 2C39 OS=Mus musculus GN=Cyp2c39 R	E=2:
2	7	SwissProt	d2::CP238_MOUSE	150	61356	9	9	4	4	0.37	Cytochrome P450 2C38 OS=Mus musculus GN=Cyp2c38 P	E=2 :
3	1	SwissProt	ef2::GRP78_MOUSE	1308	81404	55	55	21	21	2.47	78 kDa glucose-regulated protein OS=Mus musculus GN=	Ispa!
3	2	SwissProt	d2::HSP7C_MOUSE	362	78937	21	21	8	8	0.63	Heat shock cognate 71 kDa protein OS=Mus musculus GN	=Hsp.
4	1	SwissProt	ef2::CYB5_MOUSE	1217	16817	42	42	5	5	3.08	Cytochrome b5 OS=Mus musculus GN=Cyb5a PE=1 SV=2	
5	1	SwissProt	ef2::PDIA1_MOUSE	1123	64694	53	53	16	16	2.54	Protein disulfide-isomerase OS=Mus musculus GN=P4hb F	E=1 :
6	1	SwissProt	d2::CP1A2_MOUSE	1054	63034	38	38	10	10	1.31	Cytochrome P450 1A2 OS=Mus musculus GN=Cyp1a2 PE	1 SV
Z	1	SwissProt	ef2::ENPL_MOUSE	1018	103744	63	63	19	19	1.53	Endoplasmin OS=Mus musculus GN=Hsp90b1 PE=1 SV=2	
8	1	SwissProt	2::RDH7_MOUSE	1005	38455	45	45	12	12	4.07	Retinol dehydrogenase 7 OS=Mus musculus GN=Rdh7 PE	2 SV
8	2	SwissProt	@2::H17B6_MOUSE	597	38949	23	23	7	7	1.37	17-beta-hydroxysteroid dehydrogenase type 6 OS=Mus m	uscul
2	1	SwissProt	@2::MGST1_MOUSE	863	18595	25	25	3	3	2.57	Microsomal glutathione S-transferase 1 OS=Mus musculus	GN=
10	1	SwissProt	2::RL7A_MOUSE	770	35860	28	28	8	8	1.91	60S ribosomal protein L7a OS=Mus musculus GN=Rpl7a P	E=2 5
11	1	SwissProt	102::RLA0_MOUSE	763	37215	24	24	7	7	1.47	60S acidic ribosomal protein P0 OS=Mus musculus GN=Rg	Ip0 P
12	1	SwissProt	2::CP2AC_MOUSE	763	61325	35	35	14	14	2.25	Cytochrome P450 2A12 OS=Mus musculus GN=Cyp2a12 I	E=1
12	2	SwissProt	2::CP2A5_MOUSE	59	61696	5	5	2	2	0.17	Cytochrome P450 2A5 OS=Mus musculus GN=Cyp2a5 PE	2 SV
13	1	SwissProt	2::ACSL1_MOUSE	749	86078	38	38	18	18	1.90	Long-chain-fatty-acidCoA ligase 1 OS=Mus musculus GM	=Acs
13	2	SwissProt	2::ACSL5_MOUSE	297	84629	15	15	6	6	0.41	Long-chain-fatty-acidCoA ligase 5 OS=Mus musculus GM	=Acs
14	1	SwissProt	@2::RL13_MOUSE	748	28083	31	31	7	7	2.90	60S ribosomal protein L13 OS=Mus musculus GN=Rpl13 F	E=2 :
15	1	SwissProt	m2::PDIA3_MOUSE	692	64504	40	40	15	15	2.06	Protein disulfide-isomerase A3 OS=Mus musculus GN=Pdi	3 PE
16	1	SwissProt	@2::CP3AB_MOUSE	686	65154	32	32	10	10	1.25	Cytochrome P450 3A11 OS=Mus musculus GN=Cyp3a11	E=1
17	1	SwissProt	@2::UDB17_MOUSE	677	67040	34	34	9	9	0.91	UDP-glucuronosyltransferase 2B17 OS=Mus musculus GN	Ugt2
17	2	SwissProt	MZ::UD11_MOUSE	429	65361	19	19	7	7	0.80	UDP-glucuronosyltransferase 1-1 OS=Mus musculus GN=	igt1a
17	3	SwissProt	d2::UD16_MOUSE	245	65516	14	14	6	6	0.67	UDP-glucuronosyltransferase 1-6 OS=Mus musculus GN=	gt1a
18	1	SwissProt	@2::EST3A_MOUSE	668	67490	28	28	5	5	0.43	Carboxylesterase 3A OS=Mus musculus GN=Ces3a PE=1	5V=2 +
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The Report Builder tab is useful when you need a table of proteins suitable for publication. Lets assume we want to drop the 'one hit wonders' and only report proteins that have significant matches to at least 2 different peptide sequences

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4 1 SwissProt	2::CYB5 MOUSE 12	17 16817	42	42	5	5	3.08	Cytochrome b5 OS=Mus musculus GN=Cyb5a PE=1 SV=2	
5 1 SwissProt	2::PDIA1_MOUSE 11	23 64694	53	53	16	16	2.54	Protein disulfide-isomerase OS=Mus musculus GN=P4hb PE=1 1	
6 1 SwissProt d	2::CP1A2_MOUSE 10	54 63034	38	38	10	10	1.31	Cytochrome P450 1A2 OS=Mus musculus GN=Cyp1a2 PE=1 SV	
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8 1 SwissProt d	2::RDH7_MOUSE 100	05 38455	45	45	12	12	4.07	Retinol dehydrogenase 7 OS=Mus musculus GN=Rdh7 PE=2 SV	
8 2 SwissProt	2::H17B6_MOUSE 59	97 38949	23	23	7	7	1.37	17-beta-hydroxysteroid dehydrogenase type 6 OS=Mus muscul	
10 1 SwissProt	21:PIGSTI_MOUSE 80	70 35860	25	25	3	3	2.57	60S ribosomal protein L7a OS=Mus musculus GN=Pol7a DE=2	
11 1 SwissProt	2::RLAO MOUSE 70	63 37215	20	20	7	7	1.47	60S acidic ribosomal protein P0 OS=Mus musculus GN=Rpip0 P	
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13 1 SwissProt da	2::ACSL1_MOUSE 74	49 86078	38	38	18	18	1.90	Long-chain-fatty-acidCoA ligase 1 OS=Mus musculus GN=Acs	
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14 1 SwissProt	2::RL13_MOUSE 74	48 28083	31	31	7	7	2.90	60S ribosomal protein L13 OS=Mus musculus GN=Rpl13 PE=2 !	
15 1 SwissProt	2::PDIA3_MOUSE 69	92 64504	40	40	15	15	1.25	Protein disumde-isomerase As US=Mus musculus GN=Pdia3 PE	
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-	1	SwiceProt	2: PDIA1 MOUSE	1123	64694	53	53	16	16	2	54 Protein disulfide-isomerase OS=Mus musculus GN=D4hh	PE=1 SV
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14	1	SwissProt	d2::RL13_MOUSE	748	28083	31	31	7	7	2	60S ribosomal protein L13 OS=Mus musculus GN=Rpl13	PE=2 SV
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Only proteins with significant matches to at least 2 sequences remain. The filtering is very flexible, with lots of useful terms.

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2	1	SwissProt	2::CP2CT MOUSE	1337	61419	76	76	13	13	2.00	Cytochrome P450 2C29 OS=Mus musculus GN=Cyp2c29 PE=1 SV=
2	2	SwissProt	2::CP254 MOUSE	552	60887	27	27	8	8	0.88	Cytochrome P450 2C54 OS=Mus musculus GN=Cyp2c54 PE=2 SV=
2	3	SwissProt	2::CY250_MOUSE	489	61128	27	27	10	10	1.20	Cytochrome P450 2C50 OS=Mus musculus GN=Cyp2c50 PE=1 SV=
2	4	SwissProt	d2::CP2F2_MOUSE	484	59267	32	32	12	12	2.11	Cytochrome P450 2F2 OS=Mus musculus GN=Cyp2f2 PE=2 SV=1
2	5	SwissProt	2::CP237_MOUSE	339	60590	22	22	8	8	0.89	Cytochrome P450 2C37 OS=Mus musculus GN=Cyp2c37 PE=2 SV=
2	6	SwissProt	#2::CP239_MOUSE	251	60856	13	13	4	4	0.37	Cytochrome P450 2C39 OS=Mus musculus GN=Cyp2c39 PE=2 SV=
2	7	SwissProt	2::CP238_MOUSE	150	61356	9	9	4	4	0.37	Cytochrome P450 2C38 OS=Mus musculus GN=Cyp2c38 PE=2 SV=
3	1	SwissProt	d2::GRP78_MOUSE	1308	81404	55	55	21	21	2.47	78 kDa glucose-regulated protein OS=Mus musculus GN=Hspa5 PE
2	2	SwissProt	2::HSP7C_MOUSE	362	78937	21	21	8	8	0.63	Heat shock cognate 71 kDa protein OS=Mus musculus GN=Hspa8 F
4	1	SwissProt	d2::CYB5_MOUSE	1217	16817	42	42	5	5	3.08	Cytochrome b5 OS=Mus musculus GN=Cyb5a PE=1 SV=2
5	1	SwissProt	d2::PDIA1_MOUSE	1123	64694	53	53	16	16	2.54	Protein disulfide-isomerase OS=Mus musculus GN=P4hb PE=1 SV=
6	1	SwissProt	12::CP1A2_MOUSE	1054	63034	38	38	10	10	1.31	Cytochrome P450 1A2 OS=Mus musculus GN=Cyp1a2 PE=1 SV=1
Z	1	SwissProt	2::ENPL_MOUSE	1018	103744	63	63	19	19	1.53	Endoplasmin OS=Mus musculus GN=Hsp90b1 PE=1 SV=2
8	1	SwissProt	2::RDH7_MOUSE	1005	38455	45	45	12	12	4.07	Retinol dehydrogenase 7 OS=Mus musculus GN=Rdh7 PE=2 SV=1
8	2	SwissProt	2::H17B6_MOUSE	597	38949	23	23	7	7	1.37	17-beta-hydroxysteroid dehydrogenase type 6 OS=Mus musculus C
2	1	SwissProt	d2::MGST1_MOUSE	863	18595	25	25	3	3	2.57	Microsomal glutathione S-transferase 1 OS=Mus musculus GN=Mgs
10	1	SwissProt	2::RL7A_MOUSE	770	35860	28	28	8	8	1.91	60S ribosomal protein L7a OS=Mus musculus GN=Rp17a PE=2 SV=
1 H	1	SwissProt	M211RLA0_MOUSE	763	3/215	24	24			1.4/	605 acidic ribosomai protein PO OS=Mus musculus GN=Rpipo PE=1
12	1	SwissProt	M2::CP2AC_MOUSE	/03	61606	35	35	14	14	2.25	Cytochrome P450 2A12 OS=Mus musculus GN=Cyp2a12 PE=1 SV=
12	1	SwiceProt	2::0F2A5_MOUSE	740	86078	38	38	18	18	1.00	Longschain-fatty-acide-CoA ligase 1 OS=Mus musculus GN=Cyp2a5 PE=2 SV=1
13	2	SwissProt	2:ACSL5 MOUSE	297	84629	15	15	10	10	0.41	Long-chain-fatty-acidCoA ligase 5 OS=Mus musculus GN=Acil5 P
14	1	SwissProt	2::RL13 MOUSE	748	28083	31	31	7	7	2,90	60S ribosomal protein L13 OS=Mus musculus GN=Rol13 PE=2 SV=
15	1	SwissProt	2::PDIA3 MOUSE	692	64504	40	40	15	15	2.06	Protein disulfide-isomerase A3 OS=Mus musculus GN=Pdia3 PE=1 1
16	1	SwissProt	2::CP3AB_MOUSE	686	65154	32	32	10	10	1.25	Cytochrome P450 3A11 OS=Mus musculus GN=Cyp3a11 PE=1 SV=
17	1	SwissProt	2::UDB17_MOUSE	677	67040	34	34	9	9	0.91	UDP-glucuronosyltransferase 2B17 OS=Mus musculus GN=Uqt2b17
17	2	SwissProt	d2::UD11_MOUSE	429	65361	19	19	7	7	0.80	UDP-glucuronosyltransferase 1-1 OS=Mus musculus GN=Ugt1a1 PE
17	3	SwissProt	d2::UD16_MOUSE	245	65516	14	14	6	6	0.67	UDP-glucuronosyltransferase 1-6 OS=Mus musculus GN=Ugt1a6 PE
18	1	SwissProt	2::EST3A_MOUSE	668	67490	28	28	5	5	0.43	Carboxylesterase 3A OS=Mus musculus GN=Ces3a PE=1 SV=2
19	1	SwissProt	ef2::RL4_MOUSE	650	55568	34	34	11	11	1.59	60S ribosomal protein L4 OS=Mus musculus GN=Rpl4 PE=1 SV=3
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Another thing that you could easily do would be to exclude proteins from the contaminants database



The columns section of Report Manager allows you to choose which columns to include and, if required, change their order

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34 3		1	iPRG 2012	P40150	1613	66668	105	66	66	45	11.76	Heat shock protein SSB2 OS=Sacch
35 3	1	2	iPRG 2012	P11484	1590	66732	103	65	64	44	11.12	Heat shock protein SSB1 OS=Sacch
36 4		1	iPRG 2012	P10592	1591	69599	107	57	52	32	5.01	Heat shock protein SSA2 OS=Sacch
37 4		2	iPRG 2012	P10591	1161	69786	85	44	48	26	3.02	Heat shock protein SSA1 OS=Sacch
38 4	1	3	iPRG 2012	P16474	233	74479	23	8	17	6	0.32	78 kDa glucose-regulated protein hor
39 5		1	iPRG_2012	P00330	1453	37282	73	51	32	25	13.48	Alcohol dehydrogenase 1 OS=Sacch
40 5		2	iPRG_2012	P07246	101	40743	14	5	7	3	0.29	Alcohol dehydrogenase 3, mitochono
41 6		1	iPRG_2012	P00560	1382	44768	102	58	54	33	12.75	Phosphoglycerate kinase OS=Sacch
42 7		1	iPRG_2012	P00359	1361	35838	76	54	31	25	12.29	Glyceraldehyde-3-phosphate dehydro
43 7		2	iPRG_2012	P00358	1242	35938	69	48	29	24	9.89	Glyceraldehyde-3-phosphate dehydro
44 7		3	iPRG_2012	P00360	505	35842	30	20	14	12	2.47	Glyceraldehyde-3-phosphate dehydro
45 7		4	iPRG_2012	P04406	41	36201	4	2	4	2	0.21	Glyceraldehyde-3-phosphate dehydro
46 8		1	iPRG_2012	P06169	1289	61685	44	41	28	26	4.7	Pyruvate decarboxylase isozyme 1 C
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48 1	0	1	iPRG_2012	P07281	1015	15881	51	38	16	13	22.71	40S ribosomal protein S19-B OS=Sa
49 1	0 :	2	iPRG_2012	P07280	1014	15907	51	38	16	13	22.71	40S ribosomal protein S19-A OS=Sa
50 1	2	1	contaminants	P00/61	922	250/8	3/	2/	22	0	2.89	SWISS-PRUT:PUU/6111RYP_PIG Tr
51 1	2	4	IPRG_2012	P32324	784	33000	49	33	33	23	1.44	Elongation factor 2 US=Saccharomy
52 1	4	1	IPRG_2012	P10021	766	10739	20	33	10	9	95.65	Elongation factor 3A US=Saccharon
54 1	5	1	iPRG_2012	003048	700	15948	28	23	17	14	17.82	Cofilin OS=Saccharomycae caravisia
55 1	6	1	iPRG 2012	POCOV8	719	9797	42	29	15	12	207.43	40S rihosomal protein S21-A OS=Sa
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Once the list is filtered and the columns arranged as required, there is a button to export the table as CSV, which can be pasted into Excel and formatted to create a suitable figure for dropping into a publication



If you are still using Mascot 2.2 or if you have some application software that requires the results in the earlier format, and you are encountering problems with timeouts and running out of memory, here are some tips:

•Ensure you are using the Select report. If you are using a third party client that has specified Peptide summary or Protein summary, add this to the URL before opening the file: &REPTYPE=select

•Don't specify a huge number of hits 'just in case'. Choose AUTO to display all protein hits that contain at least one significant peptide match: &REPORT=AUTO

•Get rid of the yellow pop-ups: &_showpopups=FALSE

•Setting require bold red and leaving "Display non-significant matches" unchecked will minimize the number of hits: &_requireboldred=1

Mascot database search Nesult A	T O O							-			
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		_showsubsets	ShowSubSets	~		1	For a Peptide Summary, set the value to 1 to report all hits that match a subset of peptides. Default is 0 for no sub-set hits. Intermediate values set a threshold on the difference in protein score between the primary hit and the sub-set hit expressed as a fraction.				
		_requireboldred	RequireBoldRed	~		1	Set value to 1 to report Peptide Summary hits only if they contain at least one "bold red" peptide, (default 0).				
							report all matches				
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If you can't remember these URL parameters, just click on the help link

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F	ormat As Select Summary (protein hits) 🗸	Help	Help	
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MASCOT	: Very Large Searches	© 2007-2022 Matrix Science		MATRIX SCIENCE

What do we mean by Standard scoring and MudPIT scoring?

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	Standar	d protei	n score									
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		Query Observe	Mr(expt)	Mr(calc)	ppm 1	fiss Se	core Expe	et I	Rank U	Inique	Peptide	
		28 359.734	717.4537	717.4537	-0.09	0	7 4	1.2	5	U	R.LFAIVR.G	
		209 394.237	786.4596	786.4599	-0.46	0	8	13	3	0	K.LTIADVR.A	
		357 413 264	824.5139	824.5135	5.61	1	12 1	15	71 5	о 11	K. IDOULIK.C	
		715 450.736	5 899.4584	899.4588	-0.38	0	10 2	.9	2	u	K. IVDVSSDR. C	
		740 451.768	901.5217	901.5233	-1.72	0	3	24	3	U	R.VTLVDVTR.N	
		840 459.248	916.4821	916.4767	5.98	0	2	29	2	υ	K.GVEFNVPR.L	
		844 459.729	917.4452	917.4454	-0.24	0	4	15	6	υ	K.ELEETAAR.M	
		1029 473.275	944.5368	944.5331	3.97	1	3	21	3	ឋ	R.EPPSFIKK.I	
		1058 475.750	949.4864	949.4869	-0.47	0	4	22	5	U	R.SSVSLSWGK.P	
		1066 476.279	950.5433	950.5425	0.94	0	1	23	4	U	R.PLTDLQV <u>R</u> .E	
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With standard peptide summary scoring, the protein score is essentially the sum of the ions scores of all the peptides assigned to the protein. Where there are duplicate matches to the same peptide, the highest scoring match is used. A correction is applied based on the number of candidate peptides that were tested. This correction is very small unless it is a very large protein, like here, or a no-enzyme search

Despite this correction, as this example shows, when we have many low scoring matches assigned to the same protein, we can still get a high protein score, even though none of the individual peptide matches are significant



A protein with matches to just a single peptide sequence is commonly referred to as a "one-hit wonder" and is often treated as suspect. This is actually a slight oversimplification. In a search with a large number of spectra and a small database, even though the peptide false discovery rate is low, a protein can pick up multiple false matches by chance. This is easily calculated using a Poisson Distribution, where m is the average number of false matches per protein. In this example, m is 750/5268, and we would expect 650 database entries to be one-hit wonders. However, 46 entries will pick up two false matches and 2 entries will pick up three, which could mean we report 48 false proteins.

The problem isn't limited to large searches. It is the ratio between the number of spectra and the number of entries in the database that matters. So, a small search against a small database can give similar numbers

Protein Scores for MS/MS Searches												
	MudPIT p	rote	in sco	ore								
 The sum of the excess of the ions score over the identity or homology threshold for each query Plus 1 x the average threshold 												
	1249. <u>2::IP</u>	00023283	Mass: 383	2803 Score	e: 0	Ma	tches:	51(0)	Seque	ences: 4	18 (0)	
	Tax_Io	1=9606 Gene	_Symbol=TTN	Isoform 2	of Titi	n		-				
	Query	359.7341	nr(expt)	nr(calc) 717.4537	-0.09	0	Score	Expect	Rank	Unique	Peptide P.LEATUR.G	
	209	394.2371	786.4596	786.4599	-0.46	0	8	13	3	u	K.LTIADVR.A	
	334	411.2073	820.4000	820.3954	5.61	0	3	15	4	u	K. TDSGLYR. C	
	357	413.2642	824.5139	824.5135	0.48	1	12	1.1	5	U	K.RFLTLR.K	
	715	450.7365	899.4584	899.4588	-0.38	0	10	2.9	2	U	K. IVDVSSDR.C	
	740	451.7681	901.5217	901.5233	-1.72	0	3	24	3	U	R.VTLVDVTR.N	
	840	459.2484	916.4821	916.4767	5.98	0	2	29	2	υ	K.GVEFNVPR.L	
	844	459.7299	917.4452	917.4454	-0.24	0	4	15	6	U	K.ELEETAAR.M	
	1029	473.2757	944.5368	944.5331	3.97	1	3	21	3	U	R.EPPSFIKK.I	
	1058	475.7505	949.4864	949.4869	-0.47	0	4	22	5	U	R.SSVSLSWGK.P	
	1066	476.2790	950.5433	950.5425	0.94	0	1	23	4	U	R.PLTDLQV <u>R</u> .E	
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To avoid this problem, we use MudPIT protein scoring, in which the score for each peptide match is not its absolute score, but the amount that it is above the threshold. Therefore, matches with a score below the threshold do not contribute to the score. The MudPIT protein score is the sum of the score excess over threshold for each of the matching peptides plus one times the average threshold. For each peptide, the "threshold" is the homology threshold if it exists, otherwise it is the identity threshold.

So, even though a large protein like titin may pick up several random matches, with MudPIT scoring, the protein score is zero, so you don't see it listed in the report unless you specify a huge number of protein hits, as was done here to capture this screen shot.

By default, MudPIT protein scoring is used when the ratio between the number of queries and the number of database entries, (after any taxonomy filter), exceeds 0.001. This default switching point can be moved by changing the value of MudpitSwitch in mascot.dat. You can also switch between the two scoring methods by using the format controls at the top of the report.

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MASCOT	: Very Large Searches	© 2007-2022 Matrix Science		MATRIX SCIENCE

At some stage, it is likely that you will want to export the search results to another application or a relational database. If you want to write your own code, we provide a free library called Mascot Parser that provides a clean, object oriented programming interface to the result file. The supported languages are C++, Java, and Perl.

Mascot also includes a flexible export utility.

If you want the XML format, you probably know that this is what you want. If you've no idea what XML is, chances are you don't want it.

Choose CSV if you want to export to Excel - I'll show an example in a moment.

Choose pepXML if you want to export to Protein Prophet from ISB.

mzIdentML and mzTab are the standard formats from PSI for search result interchange. Mascot provides a very full implementation of mzIdentML and this is the one to choose if you are writing new application software that will use Mascot results

DTASelect is the tab separated format used by David Tabb's DTASelect program

The Mascot DAT file is the raw result file. If you need the result file for some reason, and don't have FTP or SCP access to your Mascot server, this is a convenient way to get the file.

MGF peak list is useful when you have the search result but can't find the peak list.



If you arrive here from one of the older reports, to begin with, you may need to select the required output format. Different formats have different options further down the page

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To export to Excel, simply select CSV as the format, and click on the Export Search Results button at the bottom of the page. In recent versions of Mascot, the report is prepared and then a download button is displayed. In older versions, the download would start immediately. One the download is finished, you can open it into Excel:

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Much easier and safer than "screen scraping"

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For those of you into XML, here is a sample XML file. The schema is available from our web site or your local Mascot installation.

Please read the help for details.

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XML is ideal for transferring the results to a relational database. Even Microsoft Access can open the XML file directly into database tables

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There is a very detailed help page for all of this.

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Which describes how the export script can be called from the command line or a shell prompt, as part of an automated pipeline.

I won't go into any detail here, but this means that it is possible to set up a script that will, for example, automatically convert all of your Mascot results to XML files.

Figuring out the command line arguments from the help can be tricky so, in Mascot 2.3, we added a function to display the command line corresponding to the selected options



By the way, don't delete the original result files after exporting them or your won't be able to view the standard Mascot reports in a browser.